

## Influence of Biofertilizers on Growth and Leaf Mineral Content in Peach Transplants

\*Mustafa E.A.Al-Hadethi<sup>1</sup>, Ali S.T. AL-Dulaimi<sup>2</sup> and B.M.K. Almashhadani<sup>2</sup>

1. Dep. of Horticulture, College of Agriculture, University of Baghdad, Iraq.

2. Dep. of Biology, Al-Rasheed University College.

Corresponding Author: \*Mustafa E.A.Al-Hadethi

---

**Abstract:** This study was conducted in the Department of Biology, Al-Rasheed University College- Baghdad during 2017 growing season to investigate the influence of some microbial inoculants on 1 year's old trees of "Peento" peach cultivar. The biofertilizers treatments was control treatment (B<sub>1</sub>), *Azotobacter chroococcum* (B<sub>2</sub>), *Azospirillum brasilense* (B<sub>3</sub>), *Bacillus megatherium* (B<sub>4</sub>), *Azotobacter chroococcum* + *Azospirillum brasilense* (B<sub>5</sub>), *Azotobacter chroococcum* + *Bacillus megatherium* (B<sub>6</sub>), *Azospirillum brasilense* + *Bacillus megatherium* (B<sub>7</sub>), *Azotobacter chroococcum* + *Azospirillum brasilense* + *Bacillus megatherium* to the soil (B<sub>8</sub>). The experimental design adapted in this experiment was RCBD. The number of transplant used was 24 transplants. The results indicate that the *Azotobacter chroococcum* + *Azospirillum brasilense* + *Bacillus megatherium* to the soil (B<sub>8</sub>) treatment significantly increased leaves area of 1312 cm<sup>2</sup>, leaf chlorophyll content of 34.56 mg.100g<sup>-1</sup>, increase in stem diameter of 2.24 mm and the highest average of branches length of 19.20 cm. As the same treatment has been given the highest leaf nitrogen content of 1.60 %, the highest leaf potassium content of 1.66 %, highest leaf zinc content of 18.12 ppm and highest leaf manganese content of 2.134 ppm. The lowest values for these treatments for all studied traits were in the control treatment (B<sub>1</sub>).

---

Date of Submission: 06-09-2017

Date of acceptance: 20-09-2017

---

### I. Introduction

Peaches (*Prunus persica* L.) are native to family Rosaceae. They were early cultivated in China since approximately 4000 years ago from it speeded world wide. Worldwide, peaches' production increased in the past 20 years, due to the new established orchard on large surfaces, as a result of peaches' consumption increase and also to the new technology links developed for peach culture (Iuliana, 2013). The acreage of peach in the world reached about 1499872 hectare, with production of 21083151 tons, the main producing countries are China then Italy (FAO, 2013). The estimated number of peach fruit trees in Iraq, including nearly 152273 tree produces up to 2451 tons, and the average production per tree about 16.1 kg (PCBS, 2013).

The use of microbial inoculants in agriculture has greatly increased during the past two decades (Hayat et al. 2010) as the public and private sector agricultural research and development communities work for solutions to problems associated with modern agriculture. Microbial inoculants are typically classified as biocontrol agents (also called Biopesticides) or biofertilizers (Bashan and Holguin 1998). Biofertilizers are biological products containing living microorganisms that, when applied to seed, plant surfaces, or soil, promote growth by several mechanisms such as increasing the supply of nutrients, increasing root biomass or root area, and increasing nutrient uptake capacity of the plant (Vessey, 2003). Biofertilizers can be used as complements to mineral fertilizers (Canbolat et al. 2006). Microbial inoculants mainly include free-living bacteria, fungi, and arbuscular mycorrhizal fungi (AMF) (Berg, 2009; Dodd and Ruiz-Lozano, 2012; Vessey, 2003) that were isolated from a variety of environments including soil, plants, plant residues, water, and composted manures. Many previous studies revealed biofertilizers affect some characteristics of growth and leaf mineral content (Nithya et al. 2011) found the *Azotobacter* and *Aspergillus* spp. affected significantly in most recipes vegetative growth studied in mulberry trees. (Dutt et al. 2013) found that adding five types of fungi to soil enhanced leaf mineral content (P, Zn, Mn, Fe) of apricot trees. (Al-Hadethi et al. 2014) reported that biofertilizer (Nitrobeine) increased the vegetative growth characteristics and leaf chlorophyll and nitrogen and potassium content of apricot trees. (Haggaget al. 2014) Found the biofertilizers affected significantly in most recipes vegetative growth studied in olive trees. The target of this study was to evaluate "Peento" peach cultivar parameters under biofertilizers treatments.

### II. Materials and Methods

This study was conducted in the Department of Biology, Al-Rasheed University College- Baghdad during 2017 growing season to investigate the influence of some microbial inoculants on 1 year's old trees of

“Peento” peach cultivar. Transplants were cultivated in plastic bags with a diameter of 25 cm. Transplants were healthy, similar in vigor and subjected to the same horticultural practices adapted in the region. The biofertilizers treatments were as follows:

1. The control treatment (**B<sub>1</sub>**).
2. Added the *Azotobacter chroococcum* to the soil (**B<sub>2</sub>**).
3. Added the *Azospirillum brasilense* to the soil (**B<sub>3</sub>**).
4. Added the *Bacillus megatherium* to the soil (**B<sub>4</sub>**).
5. Added the *Azotobacter chroococcum*+ *Azospirillum brasilense* to the soil (**B<sub>5</sub>**).
6. Added the *Azotobacter chroococcum*+*Bacillus megatherium* to the soil (**B<sub>6</sub>**).
7. Added the *Azospirillum brasilense* +*Bacillus megatherium* to the soil (**B<sub>7</sub>**).
8. Added the *Azotobacter chroococcum*+ *Azospirillum brasilense* +*Bacillus megatherium* to the soil (**B<sub>8</sub>**).

The experimental unit included one transplant and the number of treatment was eight and replicated three times. The experimental design adapted in this experiment was RCBD. The number of transplant used was 24 transplants. The obtained results were subjected to analysis of variance according to (Elsahookie and Wuhaib, 1990) using L.S.D 0.05 for comparing differences between various treatment means.

The following parameters were determined in this experiment:

1. Leaves area (cm<sup>2</sup>): By taken ten leaves from the middle position of the shoot randomly and measuring leaf area (cm<sup>2</sup>). By Digimizer program Windows 7 operating system, then mean of leaf area × number of leaves to calculate the total leaves area.
2. Leaf chlorophyll contents (mg.100g<sup>-1</sup> fresh weight): Representative fresh leaf sample at middle part of shoots were taken in the first week of June and used for analysis of chlorophyll were calorimetrically according to Mackinny (1941).
3. Leaf dry weight %: Various leaves were taken from the sapling was weighing then drained degree 3. While proven weight and calculated the percentage of dry matter by dividing weight after drying on weight before drying × 100.
4. Increase in stem diameter (mm): Stem diameter were measured by (Vernier) at the beginning and end of the experiment, according to the difference between them and that such an increase in stem diameter.
5. Average of branches length (cm): Taking four branches of each experimental unit at the beginning of the month of July and measured annual shoots formed during the season in each unit empirical metric tape measure and mined the average branches length.
6. Leaf Mineral Content: leaf chemical constitute: samples of twenty leaves from the middle part of shoots according to Chuntouarb and Cummings, (1981), were selected at random from each replicate (1<sup>st</sup> week of June) to measure their content of N, P, K according to Wilde et al (1985) on dry weight basis. Manganese and Zinc were determined as ppm using atomic absorption according to (Carter, 1993).

### **III. Results And Discussions**

**Effects of biofertilizers on leaves area, leaf chlorophyll content, leaf dry weight, Increase in stem diameter Average of branches length:** Data concerning the effect of treatments on leaves area, leaf chlorophyll content, leaf dry weight, increase in stem diameter and average of branches length during the experiment are listed in Table (1). The data cleared that *Azotobacter chroococcum* + *Azospirillum brasilense* + *Bacillus megatherium* (**B<sub>8</sub>**), significantly increased leaves area of 1312 cm<sup>2</sup>, leaf chlorophyll content of 34.56 mg.100g<sup>-1</sup>, increase in stem diameter of 2.24 mm and the highest average of branches length of 19.20 cm. Table (1) also shows that the treatments did not significantly affect the leaf dry weight. The lowest values for these treatments for all studied traits were in the control treatment (**B<sub>1</sub>**). The increase in vegetative traits may be attributed to bio-fertilizer, which has contributed to an increase in the rate of biochemical processes involving nitrogen, phosphorus and potassium compounds to form compounds and basic components of photosynthesis and respiration as well as their contribution to the formation and growth of a large number of enzymes (Barker and Pilbeam, 2007). As well as the effect of bio-fertilizers on the improvement of biological and physical soil properties as well as the chemical properties resulting from the release of larger quantities of nutrients available for absorption by the roots and thus influence the physiological processes such as increase the efficiency of photosynthesis in the leaves (Yu et al, 2014) and increase It produces carbohydrates and thus increases vegetative growth. These results are in agreement with those obtained by, (Kumar et al, 2013) on pear trees, (Al-Hadethi, 2015) on apricot trees; they found that the leaves area and increase in stem diameter and average of branches length positively correlated with biofertilizers in those trees.

**Table 1:** Effects of biofertilizers on leaves area, leaf chlorophyll content, leaf dry weight, Increase in stem diameter Average of branches length of Peach trees during 2017 season.

Treatments	Leaves area (cm <sup>2</sup> )	Chlorophyll content(mg.100g <sup>-1</sup> )	Leaf dry weight (%)	Increase in stem diameter (mm)	Average of branches length (cm)
B <sub>1</sub>	1122	30.38	30.65	1.66	11.26
B <sub>2</sub>	1236	32.15	30.44	1.88	14.82
B <sub>3</sub>	1188	31.22	31.19	1.80	13.19
B <sub>4</sub>	1174	31.06	30.22	1.76	13.85
B <sub>5</sub>	1244	33.50	30.56	1.98	15.92
B <sub>6</sub>	1230	32.86	30.28	1.94	17.28
B <sub>7</sub>	1272	31.14	31.12	2.02	16.77
B <sub>8</sub>	1312	34.56	31.24	2.24	19.20
<b>L.S.D 0.05</b>	<b>44.93</b>	<b>0.48</b>	<b>N.S</b>	<b>0.14</b>	<b>1.27</b>

**Effects of biofertilizers on leaf N, P, K, Zn and Mn content:** Data concerning the effect of treatments on nitrogen, phosphor, potassium, zinc and manganese are listed in Table (2). The data cleared that, *Azotobacter chroococcum* + *Azospirillum brasilense* + *Bacillus megatherium* (B<sub>8</sub>), significantly increased and gave the highest leaf nitrogen content of 1.60 %, the highest leaf phosphor content of 0.27 %, the highest leaf potassium content of 1.66%, highest leaf zinc content of 18.12 ppm and highest leaf manganese content of 2.134 ppm. Table (2) also shows the lowest values for these treatments for all studied elements were in the control treatment (B<sub>1</sub>). The result of these results is the increase in leaves area and leaf chlorophyll content as shown in Table 1, which results in the absorption of these elements to meet their vegetative needs and the growth of the tree. The addition of bio fertilizer has increased the content of peach leaves of the elements as a result of increased growth and efficiency of photosynthesis (Mosa et. al, 2016) by increasing the leaves area and leaf content of chlorophyll and increasing the soil content of these elements as a result of adding to the soil, resulting in increased absorption and increase the content of the leaves of these elements. These results are in agreement with those obtained by, (Fawzi et. al, 2010) on pear trees, (Dutt et al. 2013) on apricot trees, (Kumar et al, 2013) on pear trees; they found that the leaves mineral content positively correlated with biofertilizers in those trees.

**Table 2:** Effects of biofertilizers on leaves mineral content of Peach trees during 2017 season.

Treatments	N (%)	P (%)	K (%)	Zn (ppm)	Mn (ppm)
B <sub>1</sub>	1.09	0.16	1.26	16.19	1.745
B <sub>2</sub>	1.21	0.17	1.33	16.37	1.759
B <sub>3</sub>	1.25	0.17	1.38	16.66	1.822
B <sub>4</sub>	1.16	0.20	1.35	17.00	1.750
B <sub>5</sub>	1.55	0.19	1.48	16.78	1.818
B <sub>6</sub>	1.42	0.22	1.50	17.23	1.856
B <sub>7</sub>	1.48	0.24	1.43	17.88	1.902
B <sub>8</sub>	1.60	0.27	1.66	18.12	2.134
<b>L.S.D 0.05</b>	<b>0.13</b>	<b>0.03</b>	<b>0.16</b>	<b>0.34</b>	<b>0.040</b>

### References

- [1]. Al-Hadethi, Mustafa. E.A. 2015. Effect of Different Fertilization sources and the growth regulator (Brassinosteroids) on growth and yield of Apricot trees. Ph.D. Dissertation, Coll. of Agric., Univ. of Baghdad. pp. 153 .
- [2]. Al-Hadethi, Mustafa .E.A. , B.M.K. Almashhadani and Y.F.S. Al-Qatan. 2014. Effect of foliar application of potassium and soil biofertilizer application on the growth and yield of (Lozi) apricot cultivar (*Prunus armeniaca* L.). Zagazig J. Agric. Res., 41(5): 969-975.
- [3]. Barker, A.V and D.J, Pilbeam. 2007. Plant Nutrition. CRC Press is an imprint of Taylor & Francis Group, an Informal business.
- [4]. Bashan, Y and G, Holguin. 1998. Proposal for division of plant growth promoting rhizobacteria into two classifications: biocontrol PGPR (Plant growth-promoting bacteria) and PGPR. Soil Biol Biochem. 30:1225-1228.
- [5]. Canbolat, M, Bilen S, Çakmakçı R, Şahin F, Aydın, A. 2006. Effect of plant growth-promoting bacteria and soil compaction on barley seedling growth, nutrient uptake, soil properties and rhizosphere microflora. Biol Fertil Soils. 42:350-357.
- [6]. Carter, M.R. 1993. Soil sampling and methods of analysis, Canada Soc., Soil Sci. Lewis, London, Tokyo.
- [7]. Central Organization for Statistics and Information Technology (PCBS). The Ministry of Planning and Development Cooperation. Report production of summer fruit trees for the year 2013. Baghdad. Iraq.
- [8]. Chuntanaparb, N. and G. Cummings. 1981. Seasonal trends in concentration of nitrogen, phosphorus, potassium, calcium and magnesium in leaf portion of apple, blueberry, grape and peach, J. Am. Soc. Hort. Sci. 105 (6): 933.
- [9]. Dodd, I.C and J.M, Ruiz-Lozano .2012. Microbial enhancement of crop resource use efficiency. Curr Opin Biotechnol. 23:236-242.
- [10]. Dutt, S, S. D. Sharma and P, Kumar. 2013. Inoculation of apricot seedlings with indigenous arbuscular mycorrhizal fungi in optimum phosphorus fertilization for quality growth attributes. Journal of Plant Nutrition, 36(1):15-31.
- [11]. Elshahookie, M.M and Wuhuib, K.M. 1990. Design and Analysis of experiments. Univ. Of Bagh. Dar al hekma.
- [12]. FAO. 2012. FAO. Statistics Division 2013. Available at : ( <http://faostat.fao.org/site/339/default.aspx>) 20 July 2013.
- [13]. Fawzi, F.M., S. Elham, A. Daoud and E.A. Kandil .2010. Effect of organic and biofertilizers and magnesium sulphate on growth yield, chemical composition and fruit quality of "Le-Conte" pear trees. Nature and Science. 8 (12):273-280.
- [14]. Haggag, L.F; Merwad M.A., M.F.M. Shahin and Amira A. Fouad. 2014. Effect of NPK and bio-fertilizers as soil application on promoting growth of "Toffahi" olive seedlings under greenhouse condition. Journal of Agricultural Technology. 10(6):1607-1617.

- [15]. Hayat R, Ali S, Amara U, Khalid R, Ahmed I .2010. Soil beneficial bacteria and their role in plant growth promotion: areview. *Ann Microbiol.* 60:579–598.
- [16]. Iuliana, M.B. 2013. The impact of chemical thinning with ethrel upon the productivity of two peach varieties cultivated in Periam, Timis County. *Journal of Horticulture, Forestry and Biotechnology.* 17(2): 252- 255.
- [17]. Kumar, M.PN; R, HariomSah and Pratibha. 2013. Effect of biofertilizers on growth, yield and fruit quality in low-chill pear CV Gola. *Agric. Sci. Digest.*, 33(2): 114 – 117.
- [18]. Mackinny, G. 1941. Absorption of light by chlorophyll solutions. *J. Biol. Chem.* 140(2) 315-322.
- [19]. Mosa, W.F.A.E-G; L.S. Paszt; M. Frąç; P. Trzciński; M. Przybył; W. Treder and K. Klamkowski. 2016. The influence of biofertilization on the growth, yield and fruit quality of cv. Topaz apple trees. *Hort. Sci. (Prague).* 43(3): 105–111.
- [20]. Nithya, D, S.M. Poornima, R.Pazhanimurugan, V.Gopikrishnan, M.Radhakrishnan, D, Bhivi and R.Balagurunathan. 2011. Influence of biofertilizers and irrigation systems for the growth and yield of mulberry plants. *International Journal of Plant, Animal and Environmental Sciences.* 1(3):93-99.
- [21]. Vessey, J.K. .2003. Plant growth promoting rhizobacteria as biofertilizers. *Plant Soil.* 255:571–586.
- [22]. Wilde, S.A.; R.B.Corey, J. G. Lyer and G.K. Voigt(1985). *Soil and plant Analysis for Tree culture* 3<sup>rd</sup> Ed. Pp. 89-100. Oxford IBLT publishing Co., New Delhi.
- [23]. Yu, Xuan; Xu Liu and Tian-hui Zhu. 2014. Walnut growth and soil quality after inoculating soil containing rock phosphate with phosphate-solubilizing bacteria. *Science Asia.* 40(1): 21-27.

Mustafa E.A.Al-Hadethi. “Influence of Biofertilizers on Growth and Leaf Mineral Content in Peach Transplants.” *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, vol. 10, no. 9, 2017, pp. 90–93.