

## **Physiological and blood biochemical responses in Baladi Female Goats treated with Chromium and Selenium-E under hot summer conditions**

Eid<sup>1</sup> S. Y., El-Sayed<sup>2</sup> A. I. M., El-Zaher<sup>1</sup> H. M. and Emara<sup>1</sup> S. S.\*

<sup>1-</sup> Department of Biological Applications, Radioisotopes Applications Division, Nuclear Research Center, Atomic Energy Authority, Inshas, Cairo, Egypt, P.O.13759.

<sup>2-</sup> Department of Animal Production, Faculty of Agriculture, Benha University, Benha, Egypt, P.O.13736

\*Corresponding author: Dr. S. S. Emara

---

### **Abstract**

The present study was conducted during estrous cycle in Baladi female goats to study the effect of chromium and selenium with vitamin E on some blood metabolites to improve the productive performance of female goats under hot summer conditions. Sixty of mature Baladi does with age ranged from 2 to 3 years old and the mean body weight was 26.5±1.3 kg was used under two conditions of winter and summer seasons (30 animals/season). The animals were randomly divided into three groups (ten head in each) during both seasons, the 1st group was kept as control and the 2nd group was supplemented by chromium (chromium chloride trivalent), 0.8 mg/head/day. The 3rd group was intramuscularly injected twice a week with 2ml viteselen®, contained 0.5 mg selenium and 10.7 IU vitamin E (Se-E). Blood metabolites were assessed in the three experimental goats during the estrous cycle. The results demonstrated that serum glucose and urea concentration significantly ( $P<0.001$ ) decrease under hot season during estrous cycle, on contrary AST, TP, albumin, globulin and total cholesterol showed significantly ( $P<0.001$ ) increase under hot condition during estrous cycle phases, except estrus phase for AST, TP, globulin and albumin, and for diestrus and proestrus phase with total cholesterol, while ALT and estradiol 17  $\beta$  showed non-significant effects for season. Female's goat supplemented with Cr or Se-E showed significantly decreased ( $P<0.001$ ) in serum glucose, total cholesterol and albumin as compare with control during estrous cycle phases except estrus phase for albumin showed significantly increased, on contrast, AST, ALT and estradiol 17  $\beta$  significantly ( $P<0.001$ ) increased with Cr or Se-E supplementations as compare with control during estrous cycle phases except estrus and metestrus phase for AST and ALT, respectively, urea concentration significantly ( $P<0.001$ ) decreased with Cr or Se-E supplementations as compare with control during diestrus and estrus phases, while in metestrus phase showed significantly increase with Se-E supplementation as compared with Cr and control group. TP concentration significantly ( $P<0.001$ ) increased with Cr and significantly ( $P<0.001$ ) decreased with Se-E supplementations during estrus cycle phases, except estrus phase which showed non-significant. All of the previous parameters showed significantly interactions between treatment and season during all of estrus cycle phases except metestrus phase for glucose concentration and estrus phase for albumin concentration. It can be concluded that Cr and Se-E supplementation can enhanced the blood metabolites and estradiol 17  $\beta$  hormone concentration of female Baladi goats during estrus cycle under hot summer conditions, In order to achieve improved physiological and reproductive performance for female goats under heat stress conditions in Egypt.

**Keywords:** Chromium, Selenium-E, heat stress, blood metabolites, Goats.

---

Date of Submission: 03-02-2021

Date of Acceptance: 18-02-2021

---

### **I. Introduction**

Goats are widely found in tropical and subtropical areas as well as in temperate regions; they are bred mainly for dairy production, but also for meat and fiber [1]. Hot summer season conditions cause infertility in farm animals and appears a major source of economic loss. In response to heat stress, leaders can try out a variety of approaches to progress reproduction. These approaches usually include changing the environment (i.e. attempting to cool animals during reproduction), supplementation of antioxidants in case of semi-intensive system as goats are grazed in the open during the most of day, which protect the body defense system against excessively produced free radicals during heat stress and stabilize health status of the animal, or increasing reproductive management at heat stress periods. Heat stress affects reproduction in all major farm species [2, 3].

The great focus of chromium research was given on the incorporation between chromium and diabetes. It was as late as in the 1990s that chromium also started to be studied intensively as an essential mineral in livestock animals [4]. The major role of chromium in metabolism is promoting glucose uptake by the cells [5]. The beneficial effects of chromium can be spotted more efficiently with environmental, dietary, and hormonal stressors. In

ruminants, supplementation of Chromium is recommended during heat stress periods, while, Chromium supplementation decreases the unfavorable effects of heat stress [6].

Selenium (Se) is a perfect trace element, animated for the normal growth and animals healthy. Se has a biological mission related to vitamin E while, Se is the main component of glutathione peroxidase enzyme which involved in detoxification of hydrogen peroxide and lipid hydroperoxides. The vitamin E requirement may, therefore, be defined as the amount required to prevent peroxidation in the particular subcellular membrane which is most susceptible to peroxidation. Furthermore, Se is a main component of selenoproteins and also, involved in immune and neuropsychological function in the nutrition of animals [7]. However, a few are recognized around the effects of vitamin E supplementation on specific reproductive calamities in goats [8].

Considering, the above circumstances, the Objective of this work was to study the effect of chromium and selenium-E supplementations on some blood metabolites of does under heat stress conditions through estrous cycle phases to alleviate heat stress on native Baladi female goats in order to achieve improved physiological and reproductive performance for female goats under Egyptian conditions.

## **II. Materials and Methods**

### **1. Experimental location and ethics**

The present study was carried out in Goats Experimental Farm, Nuclear Research Center, Atomic Energy Authority, which was conducted in the desert region of Inshas, Egypt. (Latitude 31° 12' N to 22° 2' N, longitude 25° 53' E to 35° 53' E). This work was reviewed and approved by the Animal Care and Welfare committee of Egyptian Atomic Energy Authority standard operating procedures. These ethics contain relevant information on the endeavor to reduce animal suffering and adherence to best practices in veterinary care according to the international Council for Laboratory Animal Science guidelines. The experimental procedures were carried out according to the Local Experimental Animals Care committee and approved by the institutional ethics committee.

### **2. Animal and Experimental design**

The current study was carried out for two successive estrous cycles in mature female goats, while, Sixty mature female goats (30 animals/ season) aged 2-3 years old with average body weight  $26.5 \pm 1.3$  kg were randomly divided into three groups (ten head in each) during both seasons, animals in the 1st group were kept as control, the 2nd group was supplemented by chromium (chromium chloride trivalent), 0.8 mg/head/day as capsules [9] and the 3rd group was intramuscularly injected with 2 ml viteselen®, contained 0.5 mg selenium and 10.7 IU vitamin E/head/day.

### **3. Animals feeding and management**

The experimental animals were healthy and clinically free of external and internal parasites and were fed a basal ration of concentrate feed mixture (CFM) according to the allowances of **NRC**, [10] of goats. The CFM composed of 37.4% wheat bran, 27% yellow corn, 12.5% soybean meal, 10.0% undecorticated cottonseed cake, 5% rice bran, 4% sugarcane molasses, 3% limestone, 1% sodium chloride and 0.1 vitamin and minerals premix. Feed mixture was offered once daily at 9:00 a.m., based on 3.5% of body weight. Berseem hay was offered ad libitum. Fresh drinking water was always available to all animals in clean basins full of freshwater. All experimental animals were protected in semi-open yards with force shade and ventilation during day and night in summer and protection from rain in winter and kept under the same environmental and managerial conditions to the termination of the trail. The does were allowed to graze five hours daily at least.

### **4. Meteorological data**

This experiment was carried out for two successive estrous cycles, under both of mild and hot environmental conditions, air temperature (AT) and the relative humidity (RH) during day and night were recorded in the farm area, which collected from the nearest meteorological station. These data were used to calculate the daily temperature humidity index (THI), and the average of each item was calculated, temperature humidity index (THI) was calculated during mild and hot seasons according to **Marai et al.**, [11] as:  $THI = db\text{ }^{\circ}C - [(0.31 - 0.31RH) \times (db\text{ }^{\circ}C - 14.4)]$  Where, THI= temperature humidity index, db °C= dry bulb temperature in Celsius and RH = relative humidity ÷100. The values obtained are then classified as follows: <22.2 = Absence of heat stress, 22.2 to <23.3 = Moderate heat stress, 23.3 to < 25.6 = Severe heat stress and 25.6 and more = Very severe heat stress [11].

**Table 1: The values of THI during the experimental period**

Seasons	Ambient temperature°C		Relative humidity %		THI	
	Max	Mini	Max	Mini	Max	Mini
Mild	23.47 ± 0.22	14.24 ± 0.20	80.01 ± 0.78	27.44 ± 0.52	22.9	13.9
Hot	34.15 ± 0.32	23.28 ± 0.35	77.91 ± 0.62	20.30 ± 0.46	32.8	23.5

### 5. Estrus Synchronization and Blood Sampling:

All groups received 10 ml of PGF2 $\alpha$  (lutalyse) in double dose (5 mg/ dose, IM) at 11 day-intervals, followed by 500IU of hCG; then after 24 hours three fertile bucks (one buck for each group) were introduced to the does and allowed to be with does for two successive estrous cycles for estrous detection and natural mating. Blood samples were collected from the jugular vein by jugular venipuncture using disposable syringes. Serum was separated from clotted blood by centrifugation (20 min, 3000  $\times$  g) and clear serum collected and stored at -20°C until the biochemical and hormonal determinations. Samples were collected throughout different stages of estrous cycle, according to **Fatet et al., [1]**.

### 6. Biochemical and hormonal analysis

All the following parameters were determined using commercial kits manufactured by Diamond Diagnostic Company (Egypt). These parameters were total protein, albumin, total cholesterol and urea-N to indicate kidney functions. For liver function evaluation we evaluated serum concentration of aspartate amino transferase (AST) and alanin amino transferase (ALT). Serum concentrations of glucose was also determined. Globulin value was calculated by subtraction of albumin value from their corresponding total protein value. Estradiol17 $\beta$  (E2) hormones were estimated using Radioimmunoassay technique by commercial kits provided by Diagnostic product corporation, Los Angeles, USA. The unknown samples or standards are incubated with <sup>125</sup>I-RIA and antibody-coated tubes. And then, the liquid contents of the tube are aspirated and the radioactivity is determined in computerized gamma counter. All determinations were carried out in the tracer bioclimatology unit, Department of Biological Application, Nuclear research center, Atomic Energy Authority, Inshas, Cairo, Egypt.

### 7. Statistical analysis

Data were statistically analyzed using the general linear model procedure of GLM ANOVA procedure of SAS, [12]. The statistical model used was:  $Y_{ijk} = \mu + S_i + T_j + (S*T)_{ij} + e_{ijk}$ . Where, Y = the dependent variable,  $\mu$  = the overall mean,  $S_i$  = the fixed effect of season (1= mild, 2= hot),  $T_j$  = the fixed effect of treatment (1= control, 2= chromium, 3 = selenium-E),  $S*T_{ij}$  = the effect of interaction between season and treatment,  $e_{ijk}$  = random error. Significant differences between the means were verified by **Duncan [13]**.

## III. Results

### The effects of hot summer conditions and (Cr and Se-E) supplementations during estrus cycle in female goats on serum:

#### 1- Proteins

Serum total proteins concentrations of female goats during estrous cycle increased significantly ( $P < 0.0001$ ) under hot conditions during different stages of estrous cycle, except estrus phase which showed insignificant effect (Table. 2). Supplementation of does with Cr showed significant ( $P < 0.0001$ ) increase in TP during estrous cycle, except estrus phase, compared to control and Se-E. The highest TP concentration by 10.42 g/dl was at proestrus phase due to Cr supplementation versus 7.57 and 7.53 g/dl for control and Se-E, respectively.

On the other hand, treating animals with Se-E revealed fluctuated effect on TP concentration during estrous cycle. Se-E treatment significantly ( $P < 0.0001$ ) increased TP by 7.85 g/dl at diestrus phase more than control 7.05 g/dl, and showed significant decrease 6.66 g/dl at metestrus. Whereas, there is no significant difference in TP concentration at proestrus phase due to Se-E treatment compared with control. Similar trend of insignificance in TP was obtained at estrus phase due to Cr and Se-E treatments (Table. 2). Data presented in Table (2) showed a significant ( $P < 0.0001$ ) interactions between treatments and season (mild or hot conditions) in estrous cycle phases of female goats on TP concentration.

**Table 2. Serum total protein concentration of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Total Proteins (g/dl)			
	Diestrus	Proestrus	Estrus	Metestrus
Season (S)				
Mild	7.13 ± 0.10	7.24 ± 0.15	7.39 ± 0.26	7.05 ± 0.18
Hot	8.50 ± 0.28	9.77 ± 0.42	7.82 ± 0.28	8.63 ± 0.33
P- value	0.0001	0.0001	0.109	0.0001

Treatments (T)				
Control	7.05 <sup>C</sup> ±0.10	7.57 <sup>B</sup> ±0.12	7.51±0.23	7.09 <sup>B</sup> ±0.04
Cr	8.55 <sup>A</sup> ±0.40	10.42 <sup>A</sup> ±0.56	7.59±0.47	9.78 <sup>A</sup> ±0.37
Se-E	7.85 <sup>B</sup> ±0.06	7.53 <sup>B</sup> ±0.32	7.73±0.22	6.66 <sup>C</sup> ±0.17
P- value	0.0001	0.0001	0.785	0.0001
Interaction (S*T) Item				
Mild				
Control	7.03 <sup>cd</sup> ±0.11	7.44 <sup>c</sup> ±0.15	8.59 <sup>ab</sup> ±0.01	7.13 <sup>c</sup> ±0.04
Cr	6.60 <sup>d</sup> ±0.06	7.88 <sup>c</sup> ±0.01	5.73 <sup>d</sup> ±0.09	8.10 <sup>b</sup> ±0.29
Se-E	7.75 <sup>b</sup> ±0.12	6.39 <sup>d</sup> ±0.30	7.87 <sup>bc</sup> ±0.46	5.93 <sup>d</sup> ±0.19
Hot				
Control	7.07 <sup>cd</sup> ±0.18	7.69 <sup>c</sup> ±0.18	6.42 <sup>d</sup> ±0.27	7.06 <sup>c</sup> ±0.06
Cr	10.49 <sup>a</sup> ±0.36	12.96 <sup>a</sup> ±0.36	9.45 <sup>a</sup> ±0.55	11.46 <sup>a</sup> ±0.01
Se-E	7.94 <sup>b</sup> ±0.02	8.68 <sup>b</sup> ±0.30	7.58 <sup>c</sup> ±0.05	7.38 <sup>c</sup> ±0.04
P- value	0.0001	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

Female goats showed slightly higher in albumin (Alb) concentrations in hot than mild season during estrous cycle. Significant (P<0.0001) increase was recorded at proestrus and metestrus phases with mean concentrations of 3.35 and 3.36 g/dl for hot season versus 2.81 and 3.02 g/dl for mild, respectively, as shown in Table (3).

Cr and Se-E treatments showed significant (P<0.001) difference in Alb concentrations at proestrus and estrus phases compared with control. At proestrus phase, Cr and Se-E showed albumin concentrations 2.93 and 3.08 g/dl, respectively lower than control group 3.24 g/dl. Whereas, Cr and Se-E significantly increased Alb at estrus phase more than control, with concentrations of 3.26, 3.22 and 2.99 g/dl, respectively. Albumin concentrations showed significantly (P<0.01) interaction between season and treatment during estrous cycle, except estrus phase female goats (Table 3).

**Table 3. Serum albumin concentrations of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Albumin(g/dl)			
	Diestrus	Proestrus	Estrus	Metestrus
Season (S)				
Mild	3.06±0.08	2.81±0.06	3.13±0.03	3.02±0.04
Hot	2.97±0.10	3.35±0.07	3.18±0.06	3.36±0.03
P- value	0.43	0.001	0.40	0.001
Treatments (T)				
Control	3.05±0.08	3.24 <sup>A</sup> ±0.13	2.99 <sup>B</sup> ±0.04	3.21±0.06
Cr	3.12±0.16	2.93 <sup>B</sup> ±0.08	3.26 <sup>A</sup> ±0.04	3.13±0.05
Se-E	2.87±0.04	3.08 <sup>B</sup> ±0.01	3.22 <sup>A</sup> ±0.08	3.23±0.06
P- value	0.19	0.001	0.001	0.21
Interaction (S*T) Item				
Mild				
Control	3.02 <sup>ab</sup> ±0.16	2.69 <sup>c</sup> ±0.13	2.98±0.07	3.14 <sup>bc</sup> ±0.09
Cr	3.42 <sup>a</sup> ±0.05	2.64 <sup>c</sup> ±0.05	3.18±0.01	2.90 <sup>d</sup> ±0.02
Se-E	2.73 <sup>b</sup> ±0.01	3.10 <sup>b</sup> ±0.02	3.22±0.08	3.01 <sup>cd</sup> ±0.06
Hot				
Control	3.08 <sup>ab</sup> ±0.03	3.79 <sup>a</sup> ±0.01	2.99±0.02	3.29 <sup>ab</sup> ±0.06
Cr	2.82 <sup>b</sup> ±0.29	3.21 <sup>b</sup> ±0.12	3.35±0.08	3.36 <sup>a</sup> ±0.01
Se-E	2.99 <sup>ab</sup> ±0.06	3.05 <sup>b</sup> ±0.01	3.22±0.14	3.45 <sup>a</sup> ±0.03
P- value	0.01	0.01	0.539	0.014

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

**Table 4: Serum globulin concentration of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Globulin (g/dl)			
	Diestrus	Proestrus	Estrus	Metestrus
Season (S)				
Mild	4.07±0.18	4.53±0.20	4.27±0.28	4.04±0.22
Hot	5.54±0.27	6.42±0.45	4.63±0.29	5.27±0.34
P- value	0.0001	0.0001	0.225	0.0001
Treatments (T)				
Control	3.99 <sup>C</sup> ±0.21	4.48 <sup>B</sup> ±0.17	4.52±0.27	3.89 <sup>B</sup> ±0.12
Cr	5.43 <sup>A</sup> ±0.47	7.49 <sup>A</sup> ±0.53	4.33±0.48	6.65 <sup>A</sup> ±0.32
Se-E	4.98 <sup>B</sup> ±0.08	4.46 <sup>B</sup> ±0.34	4.51±0.26	3.43 <sup>C</sup> ±0.20
P- value	0.0001	0.0001	0.843	0.0001
Interaction (S*T) Item				
Mild				

<b>Control</b>	4.01 <sup>c</sup> ±0.38	5.07 <sup>b</sup> ±0.15	5.61 <sup>ab</sup> ±0.10	3.99 <sup>c</sup> ±0.19
<b>Cr</b>	3.18 <sup>d</sup> ±0.11	5.24 <sup>b</sup> ±0.04	2.55 <sup>c</sup> ±0.09	5.20 <sup>b</sup> ±0.26
<b>Se-E</b>	5.02 <sup>b</sup> ±0.15	3.29 <sup>c</sup> ±0.39	4.65 <sup>bc</sup> ±0.53	2.92 <sup>d</sup> ±0.04
<b>Hot</b>				
<b>Control</b>	3.99 <sup>c</sup> ±0.20	3.89 <sup>c</sup> ±0.18	3.44 <sup>de</sup> ±0.29	3.78 <sup>c</sup> ±0.13
<b>Cr</b>	7.68 <sup>a</sup> ±0.07	9.75 <sup>a</sup> ±0.48	6.11 <sup>a</sup> ±0.62	8.09 <sup>a</sup> ±0.01
<b>Se-E</b>	4.94 <sup>b</sup> ±0.04	5.62 <sup>b</sup> ±0.30	4.36 <sup>cd</sup> ±0.09	3.93 <sup>c</sup> ±0.08
<b>P- value</b>	0.0001	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

Averages of globulin concentrations of female Baladi goats during estrous cycle period significantly ( $P < 0.0001$ ) higher under hot season than mild season. It noticeable that, Glb at estrus phase did not significantly affected by season as shown in Table (4).

It can be observed from (Table. 4) Glb concentrations increased significantly ( $P < 0.0001$ ) in Cr treatment at estrous cycle period, except estrus phase as compared to control and Se-E groups. Proestrus phase recorded the highest concentration in Glb by 7.49 g/dl due to Cr effect, while the lowest concentration was 4.33 g/dl at estrus phase.

On the other side, Se-E treatment showed significantly ( $P < 0.0001$ ) decreased in Glb concentration as compared to control and Cr during proestrus and metestrus phases by 4.46 and 3.43 g/dl, respectively. Data presented in Table (4) showed a significant ( $P < 0.0001$ ) interactions between treatments and season (mild or hot conditions) in estrous cycle phases of female goats on Glb concentration. While, the highest Glb concentration due to Cr treatment was of 9.75 and 9.09 g/dl at proestrus and metestrus, respectively, under hot condition.

## 2- Glucose

It's obviously clear that, serum glucose concentration in does showed a significant ( $P < 0.01$ ) decrease under hot summer condition during estrus and metestrus phase of estrous cycle as compared with mild condition with means value 36.53 and 38.25 mg/dl, respectively. On contract, proestrus phase showed significant ( $P < 0.01$ ) increase in glucose concentration under hot condition in compared to mild condition with mean value 40.17 mg/dl (Table 5).

Cr and Se-E treatments showed significant ( $P < 0.0001$ ) decrease in glucose during estrous cycle phases, when compared with control (Table. 5). And also, data presented in Table (5) showed a significant ( $P < 0.0001$ ) interactions between treatments and season (mild or hot conditions) in estrous cycle phases except, metestrus phase, while, The lowest glucose concentration due to Cr was of 25.89 mg/dl at estrus phase, whereas it was about 24.87 mg/dl at diestrus due to Se-E treatment under hot condition. The highest glucose concentration due to Cr was of 42.28 mg/dl at diestrus and 45.89 mg/dl at estrus for Se-E treatment under mild condition.

**Table 5: Serum glucose concentration of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Glucose (mg/dl)			
	Diestrus	Proestrus	Estrus	Metestrus
<b>Season (S)</b>				
<b>Mild</b>	39.65 ±2.1	37.59 ±1.3	42.72 ±1.2	45.77 ±3.2
<b>Hot</b>	39.08 ±2.3	40.17 ±2.5	36.53 ±2.6	38.25 ±1.7
<b>P- value</b>	0.803	0.024	0.01	0.01
<b>Treatments (T)</b>				
<b>Control</b>	48.41 <sup>A</sup> ±2.3	53.25 <sup>A</sup> ±1.80	49.75 <sup>A</sup> ±2.3	57.63 <sup>A</sup> ±3.5
<b>Cr</b>	39.38 <sup>B</sup> ±2.9	32.33 <sup>B</sup> ±1.50	32.71 <sup>C</sup> ±1.8	33.19 <sup>B</sup> ±1.3
<b>Se-E</b>	30.29 <sup>C</sup> ±1.2	31.06 <sup>B</sup> ±0.29	36.40 <sup>B</sup> ±2.0	35.21 <sup>B</sup> ±1.0
<b>P- value</b>	0.0001	0.0001	0.0001	0.0001
<b>Interaction (S*T) Item</b>				
<b>Mild</b>				
<b>Control</b>	40.94 <sup>b</sup> ±3.0	45.66 <sup>b</sup> ±0.38	42.73 <sup>bc</sup> ±2.5	63.78 ±6.6
<b>Cr</b>	42.28 <sup>b</sup> ±5.6	35.21 <sup>c</sup> ±2.70	39.53 <sup>c</sup> ±2.2	35.91 ±2.2
<b>Se-E</b>	35.71 <sup>b</sup> ±0.7	31.89 <sup>cd</sup> ±0.48	45.89 <sup>b</sup> ±0.7	37.63 ±1.8
<b>Hot</b>				
<b>Control</b>	55.88 <sup>a</sup> ±1.6	60.83 <sup>a</sup> ±1.8	56.78 <sup>a</sup> ±2.4	51.48 ±1.6
<b>Cr</b>	36.48 <sup>b</sup> ±1.7	29.46 <sup>d</sup> ±0.41	25.89 <sup>d</sup> ±0.1	30.48 ±0.9
<b>Se-E</b>	24.87 <sup>c</sup> ±0.20	30.23 <sup>d</sup> ±0.04	26.91 <sup>d</sup> ±0.8	32.78 ±0.5
<b>P- value</b>	0.0001	0.0001	0.0001	0.402

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different

## 3- Liver function

Data presented in Table (6) about serum ALT activity showed non-significant differences according to season effect in does during estrous cycle. serum AST activity in does showed a significant ( $P < 0.01$ ) increase under hot summer condition during diestrus and metestrus phases of estrous cycle as compared with mild condition with means

value of 62.54 and 62.55 U/L, respectively. While, serum AST activity appears non-significant differences during proestrus and estrus phases according to season effect (Table 7).

**Table 6: Serum ALT activity of does during estrous cycle as affected by season, treatment and their interaction.**

Item	ALT (U/L)			
	Diestrus	Proestrus	Estrus	Metestrus
<b>Season (S)</b>				
Mild	40.60 ±0.31	40.29 ±0.37	41.99 ±0.31	37.44 ±0.6
Hot	40.37±1.04	41.28 ±0.88	42.16 ±0.55	38.34 ±0.8
<b>P- value</b>	0.8	0.072	0.696	0.112
<b>Treatments (T)</b>				
Control	37.62 <sup>B</sup> ±0.83	37.51 <sup>C</sup> ±0.83	40.65 <sup>B</sup> ±0.69	37.52 ±0.9
Cr	42.32 <sup>A</sup> ±0.56	43.62 <sup>A</sup> ±0.23	43.05 <sup>A</sup> ±0.42	38.42 ±0.4
Se-E	41.53 <sup>A</sup> ±1.10	41.25 <sup>B</sup> ±0.74	42.54 <sup>A</sup> ±0.35	37.74 ±1.1
<b>P- value</b>	0.0001	0.0001	0.0001	0.404
<b>Interaction (S*T) Item</b>				
<b>Mild</b>				
Control	39.43 <sup>b</sup> ±0.01	39.77 <sup>b</sup> ±0.33	42.94 <sup>bc</sup> ±0.64	39.78 <sup>b</sup> ±0.17
Cr	40.00 <sup>b</sup> ±0.31	43.05 <sup>a</sup> ±0.21	41.36 <sup>d</sup> ±0.31	39.99 <sup>b</sup> ±0.17
Se-E	42.38 <sup>ab</sup> ±0.61	38.08 <sup>b</sup> ±0.34	41.69 <sup>cd</sup> ±0.55	32.54 <sup>d</sup> ±0.4
<b>Hot</b>				
Control	35.81 <sup>c</sup> ±1.5	35.25 <sup>c</sup> ±1.39	38.36 <sup>c</sup> ±0.80	35.26 <sup>c</sup> ±1.5
Cr	44.64 <sup>a</sup> ±0.48	44.18 <sup>a</sup> ±0.34	44.75 <sup>a</sup> ±0.37	36.84 <sup>c</sup> ±0.5
Se-E	40.68 <sup>b</sup> ±2.1	44.43 <sup>a</sup> ±0.62	43.39 <sup>ab</sup> ±0.31	42.94 <sup>a</sup> ±0.23
<b>P- value</b>	0.0001	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

**Table 7: Serum AST activity of does during estrous cycle as affected by season, treatment and their interaction.**

Item	AST(U/L)			
	Diestrus	Proestrus	Estrus	Metestrus
<b>Season (S)</b>				
Mild	58.34 ±1.1	49.79 ±1.5	54.29 ±0.96	50.02 ±0.97
Hot	62.54 ±2.5	47.65 ±2.0	54.41 ±1.8	62.55 ±3.4
<b>P- value</b>	0.013	0.172	0.94	0.01
<b>Treatments (T)</b>				
Control	48.81 <sup>B</sup> ±1.7	37.56 <sup>B</sup> ±1.3	53.25 ±1.2	46.05 <sup>C</sup> ±0.8
Cr	65.91 <sup>A</sup> ±1.6	55.86 <sup>A</sup> ±0.9	55.51 ±1.7	57.76 <sup>B</sup> ±2.3
Se-E	66.60 <sup>A</sup> ±1.9	52.74 <sup>A</sup> ±1.9	54.29 ±2.3	65.04 <sup>A</sup> ±4.4
<b>P- value</b>	0.0001	0.0001	0.52	0.0001
<b>Interaction (S*T) Item</b>				
<b>Mild</b>				
Control	53.26 <sup>c</sup> ±2.1	42.16 <sup>c</sup> ±0.4	56.72 <sup>b</sup> ±0.63	47.36 <sup>c</sup> ±1.6
Cr	59.15 <sup>b</sup> ±1.7	52.91 <sup>b</sup> ±0.9	47.36 <sup>c</sup> ±0.94	50.02 <sup>c</sup> ±2.3
Se-E	62.62 <sup>b</sup> ±0.52	54.29 <sup>ab</sup> ±3.6	58.80 <sup>ab</sup> ±0.84	50.14 <sup>c</sup> ±0.10
<b>Hot</b>				
Control	44.36 <sup>d</sup> ±1.9	32.96 <sup>d</sup> ±1.8	49.78 <sup>c</sup> ±1.8	44.74 <sup>c</sup> ±0.11
Cr	72.67 <sup>a</sup> ±0.42	58.80 <sup>a</sup> ±1.0	63.67 <sup>a</sup> ±0.42	62.55 <sup>b</sup> ±3.3
Se-E	70.59 <sup>a</sup> ±3.5	51.18 <sup>b</sup> ±1.7	49.79 <sup>c</sup> ±4.2	79.95 <sup>a</sup> ±6.5
<b>P- value</b>	0.0001	0.001	0.001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

Cr and Se-E treatments showed significant (P<0.0001) increase in serum ALT activity during estrous cycle phases, except, metestrus phase when compared with control (Table 6). The significant (P<0.0001) interactions between season (mild or hot conditions) and treatments in estrous cycle phases are presented in (Table 3) showed highest ALT activity due to Cr treatment was of 44.75, 44.64 and 44.18 U/L at estrus, diestrus and proestrus, respectively, under hot condition, and the highest ALT activity due to Se-E treatment was of 44.43 U/L at proestrus under hot condition.

Serum AST activities were significantly (P<0.0001) higher in treated groups at diestrus, proestrus and metestrus phases than control. However, AST activity at estrus phase did not show significant differences due to Cr or Se-E treatments compared with control. AST differences between Cr and Se-E were not significant except at metestrus Cr treatment had serum AST activity about 7.28 U/L less than Se-E treatment (Table 7).

Data in Table 7 showed that, analysis of variance of obtained data revealed significant effect due to the interaction between season and treatments on AST activity during estrous cycle. While, under mild conditions, highest AST activity 62.62 U/L was recorded at diestrus due to Se-E treatment, and under hot season, highest AST activity was 79.95 U/L found at metestrus due to Se-E treatment. However, the lowest AST due to treatments was 49.79 U/L at estrus phase of Se-E treatment too.

**4- Total cholesterol**

Serum cholesterol was differed significantly ( $P<0.0001$ ) due to season at diestrus, estrus and metestrus phases. And the highest concentration of TC 3.82 mg/dl was recorded at metestrus during hot season; moreover, the lowest TC concentration was about 2.38 mg/dl during diestrus at the same season as shown in Table (8)

Female goats supplemented with Cr showed significant ( $P<0.0001$ ) decrease in TC during estrous cycle, except metestrus phase, compared with control. The lowest TC concentration of 2.35 mg/dl was found due to Cr at estrus phase. Se-E treatment had the same trend of decrement at estrous cycle, but at metestrus Se-E increased ( $P<0.0001$ ) in TC 3.88 mg/dl as compared with control 2.89 mg/dl. The lowest cholesterol concentration due to Se-E was about 2.66 mg/dl at estrus and diestrus phases (Table 8). Data presented in table (8) showed a significant ( $P<0.0001$ ) interactions between treatments and season (mild or hot conditions) in estrous cycle phases of female goats on total cholesterol.

**Table 8: Serum total cholesterol concentration of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Total cholesterol (mg/dl)			
	Diestrus	Proestrus	Estrus	Metestrus
<b>Season (S)</b>				
Mild	3.50 ±0.15	3.05 ±0.08	2.40 ±0.05	2.69 ±0.11
Hot	2.38 ±0.16	2.85 ±0.13	3.10 ±0.15	3.82 ±0.16
P- value	0.0001	0.099	0.0001	0.0001
<b>Treatments (T)</b>				
Control	3.37 <sup>A</sup> ±0.22	3.19 <sup>A</sup> ±0.16	3.24 <sup>A</sup> ±0.21	2.89 <sup>B</sup> ±0.19
Cr	2.79 <sup>B</sup> ±0.23	2.84 <sup>B</sup> ±0.14	2.35 <sup>B</sup> ±0.07	2.99 <sup>B</sup> ±0.14
Se-E	2.66 <sup>B</sup> ±0.21	2.82 <sup>B</sup> ±0.08	2.66 <sup>B</sup> ±0.05	3.88 <sup>A</sup> ±0.21
P- value	0.012	0.018	0.0001	0.0001
<b>Interaction (S*T) Item</b>				
Mild				
Control	3.42 <sup>a</sup> ±0.34	2.73 <sup>c</sup> ±0.14	2.41 <sup>bc</sup> ±0.07	2.20 <sup>d</sup> ±0.12
Cr	3.66 <sup>a</sup> ±0.22	3.39 <sup>ab</sup> ±0.15	2.20 <sup>c</sup> ±0.02	2.98 <sup>c</sup> ±0.25
Se-E	3.43 <sup>a</sup> ±0.26	3.02 <sup>bc</sup> ±0.06	2.61 <sup>b</sup> ±0.10	2.88 <sup>c</sup> ±0.01
Hot				
Control	3.31 <sup>a</sup> ±0.29	3.65 <sup>a</sup> ±0.23	4.08 <sup>a</sup> ±0.23	3.58 <sup>b</sup> ±0.22
Cr	1.93 <sup>b</sup> ±0.18	2.29 <sup>d</sup> ±0.08	2.51 <sup>bc</sup> ±0.14	3.01 <sup>c</sup> ±0.09
Se-E	1.89 <sup>b</sup> ±0.05	2.62 <sup>cd</sup> ±0.11	2.72 <sup>b</sup> ±0.01	4.88 <sup>a</sup> ±0.05
P- value	0.002	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

**5- Urea-N**

As shown in Table 9 urea concentrations were significantly ( $P<0.0001$ ) higher during estrous cycle at mild than hot season. Serum urea gradually increased in mild season from diestrus phase to reach highest concentration 77.59 mg/dl at estrus phase then decreased to lowest concentration 55.83 mg/dl at metestrus. Whereas, does exhibited fluctuated trend of urea during estrous cycle in hot season, recorded the lowest concentration 41.4 mg/dl at proestrus phase and the highest concentration 63.89 mg/dl at estrus phase. Data in Table 9 showed that female goats treated with Cr revealed significant ( $P<0.0001$ ) decrease in serum urea at diestrus and proestrus phases, the latest had marked urea decrease 30.69 mg/dl as compared with control. Se-E group had significant ( $P<0.0001$ ) decrease in urea during estrous cycle, except proestrus, in compared to control. The lowest urea concentration showed due to Se-E treatment by 35.73 mg/dl at metestrus phase. Urea concentrations showed significantly ( $P<0.0001$ ) interaction between season and treatment during estrous cycle in female goats (Table 9).

**Table 9: Serum urea concentration of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Urea (mg/dl)			
	Diestrus	Proestrus	Estrus	Metestrus
<b>Season (S)</b>				
Mild	60.59 ±2.4	73.19 ±5.5	77.59 ±3.2	55.83 ±3.7
Hot	50.24 ±2.1	41.40 ±2.6	63.89 ±3.4	46.08 ±2.1
P- value	0.0001	0.0001	0.0001	0.0001
<b>Treatments (T)</b>				
Control	66.08 <sup>A</sup> ±3.0	71.11 <sup>A</sup> ±2.9	78.05 <sup>A</sup> ±5.6	43.65 <sup>B</sup> ±0.61
Cr	59.27 <sup>B</sup> ±1.2	30.69 <sup>B</sup> ±0.37	77.31 <sup>A</sup> ±2.5	73.48 <sup>A</sup> ±3.10
Se-E	40.89 <sup>C</sup> ±1.5	70.11 <sup>A</sup> ±7.8	56.86 <sup>B</sup> ±2.4	35.73 <sup>C</sup> ±0.89
P- value	0.0001	0.0001	0.0001	0.0001
<b>Interaction (S*T) Item</b>				
Mild				
Control	77.72 <sup>a</sup> ±2.9	81.53 <sup>b</sup> ±3.6	102.99 <sup>a</sup> ±1.5	41.31 <sup>d</sup> ±0.65

Cr	57.34 <sup>bc</sup> ±1.6	31.28 <sup>d</sup> ±0.73	67.17 <sup>c</sup> ±1.0	86.51 <sup>a</sup> ±1.90
Se-E	46.74 <sup>d</sup> ±0.65	106.79 <sup>a</sup> ±1.2	62.64 <sup>c</sup> ±2.7	39.68 <sup>d</sup> ±0.33
Hot				
Control	54.44 <sup>c</sup> ±0.20	60.68 <sup>c</sup> ±1.7	53.11 <sup>d</sup> ±4.2	45.99 <sup>c</sup> ±0.35
Cr	61.21 <sup>b</sup> ±1.70	30.11 <sup>d</sup> ±0.05	87.46 <sup>b</sup> ±2.7	60.46 <sup>b</sup> ±2.40
Se-E	35.05 <sup>c</sup> ±1.8	33.42 <sup>d</sup> ±3.2	51.09 <sup>d</sup> ±3.4	31.79 <sup>e</sup> ±0.63
P- value	0.0001	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

#### 6- Estradiol-17β

Data in Table (10) showed there was no significant difference in serum estradiol-17β concentrations of female goats as affected by season, during estrous cycle, except at diestrus while, E2 concentration was higher in hot 29.20 pg/ml than mild season 22.80 pg/ml; also, E2 concentrations were insignificantly higher at estrus and metestrus phases in hot than mild season. E2 showed the regular traditional pattern of increase and decrease during estrous phases, while, the highest level was showed estrus phase with mean values of 45.97 and 48.44 pg/ml for mild and hot season, respectively (Table 10).

Analysis of variance for obtained data revealed highly significant (P<0.0001) effect due to treatments on serum E2. In general, treatment groups showed E2 concentrations higher than control during estrous period. Se-E treatment showed significantly increased in E2 concentrations during estrous cycle more than Cr and control groups. The highest level was about 62.49 pg/ml for Se-E versus 35.02 and 44.11 pg/ml for control and Cr, respectively during estrus phase. In addition, treatment of Cr showed significant increase in E2 concentration at diestrus and estrus phases more than control with values of 25.26 and 44.11 pg/ml versus 21.61 and 35.02 pg/ml, respectively for control group (Table 10). E2 concentrations showed significantly (P<0.0001) interaction between season and treatment during estrous cycle in female goats (Table 10).

**Table 10: Serum Estradiol-17β concentration of does during estrous cycle as affected by season, treatment and their interaction.**

Item	Estradiol-17β (pg/ml)			
	Diestrus	Proestrus	Estrus	Metestrus
Season (S)				
Mild	22.80 ±1.50	30.45 ±3.9	45.97 ±4.2	25.48 ±2.9
Hot	29.20 ±0.72	29.71 ±1.1	48.44 ±1.6	27.21 ±1.5
P- value	0.0001	0.74	0.348	0.426
Treatments (T)				
Control	21.61 <sup>C</sup> ±1.2	21.98 <sup>B</sup> ±1.9	35.02 <sup>C</sup> ±2.4	19.87 <sup>B</sup> ±2.5
Cr	25.26 <sup>B</sup> ±1.7	22.18 <sup>B</sup> ±1.5	44.11 <sup>B</sup> ±2.3	23.63 <sup>B</sup> ±1.5
Se-E	31.25 <sup>A</sup> ± 1.0	46.07 <sup>A</sup> ±4.0	62.49 <sup>A</sup> ±4.2	35.53 <sup>A</sup> ±3.2
P- value	0.0001	0.0001	0.0001	0.0001
Interaction (S*T) Item				
Mild				
Control	17.95 <sup>c</sup> ± 1.6	14.01 <sup>c</sup> ±1.2	27.22 <sup>d</sup> ±1.9	12.89 <sup>c</sup> ±1.9
Cr	17.36 <sup>c</sup> ±0.01	17.57 <sup>c</sup> ±0.8	36.23 <sup>cd</sup> ±3.3	17.39 <sup>c</sup> ±0.5
Se-E	33.27 <sup>a</sup> ±1.8	59.77 <sup>a</sup> ±4.5	74.46 <sup>a</sup> ±6.1	46.16 <sup>a</sup> ±4.2
Hot				
Control	25.28 <sup>c</sup> ±1.2	29.96 <sup>b</sup> ±1.3	42.82 <sup>bc</sup> ±3.2	26.84 <sup>b</sup> ±3.8
Cr	33.16 <sup>a</sup> ±0.83	26.79 <sup>b</sup> ±2.2	51.98 <sup>b</sup> ±0.5	29.87 <sup>b</sup> ±1.5
Se-E	29.24 <sup>b</sup> ±0.32	32.36 <sup>b</sup> ±1.8	50.53 <sup>b</sup> ±3.0	24.91 <sup>b</sup> ±1.9
P- value	0.0001	0.0001	0.0001	0.0001

Means with different letters (A, B and C or a, b, c,...) in the same column are significantly different.

## IV. Discussion

**The effects of hot summer conditions and (Cr and/ or Se-E) supplementations during estrus cycle in female goats on:**

### 1- Proteins

In harmony with our results plasma albumin in blood of buffaloes were significantly higher during summer, while plasma total protein and globulin were significantly lower when compared with values of winter season showed by **Chaudhry et al., (1995)**. And, **Kamal, [14]** reported that during summer an extension in plasma volume occurred beside an increase in plasma total protein to match colloid osmotic pressure. The increase in plasma globulin noticed in summer is thus a trail to increase the plasma TP in order to maintain the osmotic pressure.

In disagreement of our results **El-Gaafarawy et al., [15]** found that high ambient temperature during summer, accompanied with a decrease in plasma total protein, albumin and globulin. And also, **Dangi et al., [16]** showed a significant decrease in total protein concentration in goats during heat stress. While, plasma total protein decreased from 6.56 to 5.88 g/dl in Baladi goats exposed to short-term heat stress. This may be due to



increase in plasma volume as a result of heat shock which causes decreases in plasma protein concentration [17].

Previous studies found insignificant changes in globulin concentrations due to season of the year [18]. In this respect, the current results disagreed with the later authors since the present results revealed an increase in globulin under hot than mild season during estrus cycle. The present results of Cr supplementation are in harmony with the findings of **El-Masry et al.**, [19] who recorded a significantly increased in serum total proteins in calves received 0.6 mg Cr/kg DM under heat stress conditions when compared with non-Cr treated calves. The increase of TP in response to Cr supplementation may be attributed to nitrogen absorption improvement [20], increased amino acid synthesis in the liver in which Cr may enhance amino acid synthesis possibly via insulin and incorporation of several amino acids into protein [21]. In the same trend, **Wang et al.**, [22] revealed that supplementation of pigs with Cr resulted in significant increase in serum TP. In disagreement of our results **El-Masry et al.**, [19] stated that supplementation calves with Cr under heat stress conditions insignificantly increased Alb concentration as compared to untreated animals.

Our results about Cr treatment are in harmony with the findings of **El-Masry et al.**, [19] in calves exposed to heat stress, while, there was significant increase in globulin concentrations in Cr treated animals more than untreated. This increase may be attributed to an improvement in immune response in treated calves, since Cr may have an effect on certain enzymes that increase immunoglobulin synthesis [21]. Also, **Soltan et al.**, [23] found that supplementation of heat stressed calves with 3 mg Cr/head/day significantly increased blood globulin concentration as compared with control group.

On contrary, the current results about globulin concentration disagreed with the finding of **Al-Saiady et al.**, [24] who reported that treating heat stressed Holstein cows with Cr resulted in significant decrease in globulin concentration during lactation period as compared to control group.

Results in the current study of Se-E treatment in accordance with **Krajnicakova et al.**, [25] who found a significant increase in TP on day 40 postpartum in goats. Also, treatment Baladi goats with Se-E caused a remarkable increased in TP concentration in all treated animals in early, mid- and late-pregnant and lactating goats when compared to control group under southern Sinai conditions [26]. **Soliman et al.**, [27] reported that ewes injected with Se-E at late-gestation and during suckling period significantly increased plasma total protein comparing with control ewes. Our results disagreement with **Kumar et al.**, [28] who stated that supplementation of sheep with Se (0.15mg Se/kg of diet) through sodium selenite had no effect on serum TP.

The decrease in albumin concentrations during diestrus and proestrus phases due to Se-E treatment of the current study agreed with the findings of **Esa**, [26] who indicated that albumin decreased in Se-E treated female goats at pre-mating stage than control group under Southern Sinai conditions. Moreover, **El-Shahat and Abd El-Monem**, [29] reported that Baladi ewes supplemented with Se-E, 2 weeks before mating and extended through pregnancy till lambing showed a significant increase in TP. They also noticed that higher concentration of Alb was found in vitamin E alone supplemented-ewes compared to vitamin E plus Se or Se alone supplemented ewes, however, ewes received Se alone had significantly lower serum TP, Alb and globulin.

On the other hand, the significant increases in globulin of Se-E treated goats were conformed to the findings of **Mahmoud et al.**, [30] who reported an improvement in blood metabolites in sheep administrated with selenium. This result may be due to improvement of protein anabolism, decrease of protein catabolism. In addition, the increase in the other blood metabolites could be ascribed to the improvement of feed efficiency by vitamin E and Se injection that improve by the way the overall animal health and/or reproductive performance.

Se-E supplemented Baladi ewes group had significantly higher total globulins than control group and the major changes in total globulins fractions was in the  $\gamma$ -globulins. The  $\gamma$ -globulins concentrations were significantly higher in Se-E supplemented group compared with Se group control. The authors attributed this behavior to the fact that animals exposed to various antigenic agents, resulting in an increase in the IgG production [31].

## 2- Glucose

In the current study, results of Cr are in harmony with those of [32] on stressed calves; [33] in early lactation cows and [19] in solar radiation exposed calves. These authors stated that animals treated with Cr had serum glucose concentration less than untreated animals. The decrease in glucose concentration may be attributed to the depression in cortisol level which has been found to be associated with reducing gluconeogenesis process and increasing glucose utilization as a response to an increase in insulin concentrations, since Cr seems to be an integral component of glucose tolerance factor to potentiate the action of insulin, [34].

On the contrary, **Nikkhah et al.**, [35] in heat-stressed dairy cows found that serum concentrations of glucose did not affected by treatment with Cr. In addition, **Malik et al.**, [36] in a study of Cr on blood glucose and liver enzymes in rabbits, stated an increase in blood glucose level at dose 200  $\mu$ g chromium chloride and further increase in blood glucose observed at dose 400  $\mu$ g.

The current results of glucose decrement due to Se-E injection are disagreement with the findings of **Mahmoud et al.**, [30] who reported that rams injected with a combination of Se-E showed significant increase in serum glucose as compared with the control group. In the same line results obtained by **Tahmasbi et al.**, [37]

in heat stressed lactating cows showed that glucose for selenium-vitamin E group was higher than untreated group. **Mahmoud et al.**, [30] attributed the increase in serum glucose may be due to improvement of protein anabolism and decreased protein catabolism.

### 3- Liver function

Concerning the levels of liver enzymes, some studies showed that serum AST activity was higher in summer, while serum ALT was insignificantly affected by season in Barki and Rahmani ewes [38], Also, the significant increase in AST and ALT activities in cows under hot condition was recorded by **El- Masry et al.**, [39].

The current results showed an increase in ALT due to hot season in a harmony with results of **Sharma and Kataria**, [40] who recorded an increase in serum ALT activity during heat stress period in goats. The increase in activities of serum AST and ALT in the heat stressed animals may be due to the increase in stimulation of gluconeogenesis corticoids under hot conditions. However, the increase in AST and ALT may be due to the impaired function of the liver under heat stress [41].

The current results are in agreement with **Malik et al.**, [36] who reported that serum ALT activity significantly increased in rabbits groups which received either 200 or 400 µg CrCl<sub>3</sub> for 60 days when compared with control group. On the other side, the increase in serum ALT activity during estrous cycle in goats supplemented with Cr was disagreed with the findings of **Paul et al.**, [42] who revealed significant decrease in ALT activity in the Cr supplemented bucks as compared with control group in Black Bengal bucks. Furthermore, **Wang et al.**, [22] indicated insignificantly decreased in ALT activity in supplementation pigs with different forms of Cr.

In agreement of our results serum AST activity was increased significantly under the effect of Cr supplementation in compared to control group [36]. Nevertheless, these results agreed with the findings of **Yazaki et al.**, [43]; **Al-Bandr et al.**, [44] who reported increase in serum AST activity due to Cr supplementation. On the other hand, the increase in AST activity disagreement with **Wang et al.**, [22] who reported that supplementation of pigs with different forms of Cr insignificantly decreased AST activity.

The present increase in serum ALT activity due to Se-E treatment are in accordance with **Shashidhar and Prasad**, [45] who found an increase in ALT activity in adult goats supplemented with Se. and also, **Singh et al.**, [46] reported ALT activity increased in buffalo calves supplemented with Se. On the contrary, our results disagreed with **Saleh**, [47] in pregnant ewes and **Esa**, [26] in pregnant and early lactating goats under southern Sinai conditions, who reported a decrease in serum ALT activity in animals treated with Se-E during the previously mentioned reproductive periods compared to untreated group.

The current Results of Se-E supplementation are disagreement with **Esa**, [26] who reported a significant decrease in serum AST activity in female goats after treatment with Se-E comparing with untreated goats. Also, **Saleh**, [47] reported that injection with Se-E decreased AST at parturition and early lactation in ewes. Many authors stated that protective dose of vitamin-E of pregnant ewes prevent hepatic damage and decrease AST activity [48, 49] [50]. Furthermore, **Kumar et al.**, [28] reported that the activity of AST was not affected due to supplementation of Se either through inorganic or organic source in the diet of lambs.

### 4- Total cholesterol

Our results about total cholesterol increase during estrus and metestrus under hot conditions agreement with **Alameen and Abdelatif**, [51] who found that serum levels of cholesterol and triglycerides were higher during summer compared to respective winter values. This is probably related to the role of the compound in ovary steroidogenesis, so that the total cholesterol concentrations are under control of the complex of factors [52].

Otherwise, the decrease in total cholesterol concentration with the increase in environmental temperature during diestrus and proestrus in our results are in harmony with **Marai et al.**, [53] who attributed the marked decrease in cholesterol concentration under heat stress to dilution as a result to the increase in total body water or to the decrease in acetate concentration, which is the primary precursor for the synthesis of cholesterol. Furthermore, the marked increase in glucocorticoid hormone level in heat stressed animals may be another factor causing the decline in blood cholesterol.

And also, **Watson et al.**, [54] explained the significantly lower serum cholesterol levels in summer compared to winter values is probably related to metabolic disorder associated with exposure of cows to hot environment which may accelerate body fat catabolism. Furthermore, **Ocak and Okan**, [55]; **Pandey et al.**, [56] found significant decline in the cholesterol during the summer season in goats and Marwari goats, respectively. While, the decreased cholesterol concentration during the heat stress may attributed to lowered thyroid activity [56] or decreased feed intake during hot summer and consequent reduction in intake of dietary cholesterol [57].

Concerning results of chromium treatment that revealed significant decrease in serum total cholesterol during estrus cycle, which clearly showed during hot season are in agreement with the findings of **Wang et al.**, [22] who suggested that supplementation with different forms of Cr significantly decreased total cholesterol in pigs. In

addition, **El-Masry et al.**, [19] found that serum cholesterol decreased significantly in calves received 0.6 mg Cr/kg DM under heat stress conditions when compared with non-Cr treated calves.

The present results of mild season which recorded an increase in serum total cholesterol during most of estrous cycle phases disagreed with the findings of **Bunting et al.**, [58] who showed that Cr supplementation decreased total cholesterol under moderate environmental conditions. The previous studies of **Depew et al.**, [59] revealed that Cr supplementation has no effect on concentration of serum total cholesterol and triglycerides.

The decrease of serum total cholesterol due to Se-E treatment especially at hot season condition during estrus cycle are in harmony with results of **Brzoska and Brzoska**, [60] who found that cows received Se in the form of sodium selenite showed significant decrease in plasma total cholesterol. On the other side, the current Se-E results disagreed with those of **Kalmath et al.**, [61] who revealed significant increase in serum total cholesterol during summer compared to winter season in both control and Se-E supplemented Hallikar cattle groups. The authors claimed that the higher concentrations of cholesterol during summer season might support the enhanced cortisol synthesis that occurs during summer stress as the cholesterol acts as a precursor for the synthesis of steroid hormones in the body.

Also, obtained results disagreed with **Kumar et al.**, [62] in Beetal goats, **Sejian et al.**, [63] in Malpura ewes and **Das et al.**, [64] in buffaloes and his increase in circulating cholesterol may be to support hepatic gluconeogenesis during adaptive mechanisms [63]. From another view, the variation in cholesterol level may be accompanied with the changes in cortisol concentration, since the change in cholesterol concentration is controlled by adrenocorticotrophic hormone, which has an effect on some steps that related to the conversion of cholesterol to pregnanolone and subsequently to cortisol [65].

## 5- Urea-N

In the present study, serum urea concentrations during hot season were lower than mild at estrous cycle, and in harmony with our results **Dixon et al.**, [66] reported that hot environment reduced nitrogen balance in Merino x Border Leicester sheep, probably due to the decrease of total dry matter intake and increase of panting. The depression in blood urea-N associated with heat stress in animals may be due to more reabsorption of urea-N from the blood to the rumen to compensate the decrease in rumen ammonia-N according to the decrease in feed intake and digestible nitrogen consumption [67]. In addition, the increase in urinary nitrogen excretion under severe heat stress conditions as indicated by a negative nitrogen balance may also contribute to the decrease of serum urea level under such conditions [68].

On the contrary, **Shwartz et al.**, [69] found that heat stress increased plasma urea-N concentration in lactating Holstein cows. Particularly, **Momtmurro et al.**, [70] suggested that the high level of urea was due to the low energy/protein ratio and to gluconeogenesis by protein degradation in conditions of insufficient energy for growth, and indicated that urea is normally higher when protein is in excess in the diet or there is a low energy / protein ratio.

Concerning treatment effect, applying Cr to goats showed a decrease of serum urea-N concentrations during most estrus cycle phases as compared with control. In agreement with our results **Wang et al.**, [22] found that supplementation of pigs with Cr in different forms decreased serum concentration of urea. Also, **Yanchev et al.**, [71] suggested that supplemental Cr may have caused reduced diet consumption, less feed protein intake, lower level of released NH<sub>3</sub> and better NH<sub>3</sub> utilization by rumen microorganisms, thus reducing the level of ammonium that has to be detoxicated as urea-N in the liver.

In accordance of the present results of Se-E supplementation, **Esa**, [26] reported that supplemented pre-mating and pregnant Baladi goats with Se-E showed significant decrease in serum urea as compared to untreated group. Contrarily, **El-Shahat and Abdel Monem**, [29] stated that Se-E treated Egyptian Baladi ewes under subtropical conditions not reveal significant difference in serum urea-N concentration among different groups.

## 6- Estradiol-17 $\beta$

Our results about non-significant effects of season on estradiol-17 $\beta$  concentration in female goats are disagreement with **Ozawa et al.**, [72] who found that heat stress reduces plasma concentrations of estradiol-17 $\beta$  and lower follicular estradiol concentration, and also, delay ovulation, while, the mechanisms by which heat stress alters the concentrations of circulating reproductive hormones are not known. Some effects of heat stress may involve adrenocorticotrophic hormone (ACTH) and increase cortisol secretion. ACTH has been reported to block estradiol-induced sexual behavior [73]. Heat stress during follicular recruitment suppresses subsequent growth to ovulation, accompanied by decreased LH receptor level and estradiol synthesis in the follicles [72] [74].

The increase of estradiol-17 $\beta$  throughout estrous cycle due to Crcl3 supplementation in the present study agreed with the findings of **Balicka et al.**, [75] in dairy heifers, who found that heifers fed Cr in different doses developed Graffian follicles and estrus incidence, and this increase in E2 concentrations during estrus may reflect its enhancing effect on Graffian follicles growth and maturation and subsequently released amount of estradiol-17 $\beta$  from granulosa cells. Increased corticosteroid secretion has been suggested while, it can be inhibit

GnRH and thus LH secretion [76]. Many studies confirmed the decrease of cortisol due to Cr supplementations which in turn may act on alleviate the adverse effects of heat stress on reproductive hormones [77]. Furthermore, Tuormaa, [78] stated that chromium exerts a significant influence on follicular maturation and luteinizing hormone.

In accordance of the effect of Se-E supplementation on estradiol 17  $\beta$  Prasdini, et al., [79] showed that Se-E supplementation provides a real strong influence in increase concentrations of estradiol 17  $\beta$  during estrus in dairy cows compared with control group. The supplementation of Se-E to repeat breeder and an oestrous buffaloes relieved the oxidative stress as shown by reduced the levels of lipid peroxidation and superoxide dismutase activity and finally, improved blood biochemical composition in Se-E supplemented animals [80].

## V. Conclusion

It can be concluded that Cr and Se-E supplementation can enhanced the blood metabolites and estradiol 17  $\beta$  hormone concentration of female Baladi goats during estrus cycle under hot summer conditions, In order to achieve improved physiological and reproductive performance for female goats under heat stress conditions in Egypt.

### Conflict of Interest

The authors declare that they have no conflict of interest.

## References

- [1]. Fatet, A., M.-T. Pellicer-Rubio, and B.J.A.r.s. Leboeuf, *Reproductive cycle of goats*. 2011. **124**(3-4): p. 211-219.
- [2]. Ahmad, S. and M. Tariq, *Heat stress management in water buffaloes: a review*. J Revista Veterinaria, 2010. **21**(1).
- [3]. Sivakumar, A., G. Singh, and V. Varshney, *Antioxidants supplementation on acid base balance during heat stress in goats*. J Asian-Australasian Journal of Animal Sciences, 2010. **23**(11): p. 1462-1468.
- [4]. Amata, I., *Chromium in livestock nutrition: A review*. J Global Advanced Research Journal of Agricultural Science, 2013. **2**(12): p. 289-306.
- [5]. Davis, C.M. and J.B. Vincent, *Isolation and characterization of a biologically active chromium oligopeptide from bovine liver*. J Archives of Biochemistry Biophysics, 1997. **339**(2): p. 335-343.
- [6]. Şahin, K., O. Küçük, and N. Şahin, *Effects of dietary chromium picolinate supplementation on performance and plasma concentrations of insulin and corticosterone in laying hens under low ambient temperature*. J Journal of animal physiology animal nutrition 2001. **85**(5- 6): p. 142-147.
- [7]. Meschy, F., *Recent progress in the assessment of mineral requirements of goats*. J Livestock Production Science, 2000. **64**(1): p. 9-14.
- [8]. Koyuncu, M. and H. Yerlikaya, *Short Communication effect of selenium-vitamin E injections of ewes on reproduction and growth of their lambs*. J South African Journal of Animal Science, 2007. **37**(4): p. 233-236.
- [9]. Williams, J., et al., *Influence of yeast culture, chromium, and thermal challenge on N and mineral balance in lambs*. J J. Anim. Sci, 1994. **72**(Suppl 2): p. 86.
- [10]. Council, N.R., et al., *Nutrient requirements of small ruminants: sheep, goats, cervids, and new world camelids*. 2007.
- [11]. Marai, I., *Fattening performance, some behavioural traits and physiological reactions of male lambs fed concentrates mixture alone with or without natural clay, under hot summer of Egypt*. J Annals of arid zone, 2000. **39**: p. 449-460.
- [12]. SAS, *SAS Institute Inc*, ed. U.s. Guide. Vol. v9.0. 2009, Cary, NC, USA: Statistical Analysis System Institute Inc.
- [13]. Duncan, D.B., *Multiple range and multiple F tests*. Biometrics, 1955. **11**(1): p. 1-42.
- [14]. Kamal, T. *Water turnover rate and total body water affected by different physiological factors under Egyptian environmental conditions*. in *Panel proceedings series (IAEA)*. 1982.
- [15]. El-Gaafarawy, A.M.S.E.-S.E.R.E.-S.a.S.A. and Ibraheem., *Effect of seasonal variations on some physiological parameters and blood biochemical changes of baladi cattle in middle Egypt*. . Egyptian J. Appl. Sci., 2004. **19**: p. 27 – 34.
- [16]. Dangi, S.S., et al., *Expression profile of HSP genes during different seasons in goats (Capra hircus)*. J Tropical animal health production, 2012. **44**(8): p. 1905-1912.
- [17]. Helal, A., et al., *Effect of heat stress on coat characteristics and physiological responses of Balady and Damascus goats in Sinai, Egypt*. J American-Eurasian Journal of Agricultural Environmental Science 2010. **7**(1): p. 60-69.
- [18]. Okoruwa, M., *Effect of heat stress on thermoregulatory, live bodyweight and physiological responses of dwarf goats in southern Nigeria*. J European Scientific Journal, 2014. **10**(27): p. 255-264.
- [19]. El-Masry, K.A., El-Fouly, H. A. and Gabr, S. A., *Physiological action of chromium on some blood biochemical constituents and immune response in relation to growth performance of heat stressed calves*. . Egyptian J. Nutrition and Feeds, , 2001. **4** (special issue): p. 453-463.
- [20]. Kornegay, E., et al., *Supplemental chromium picolinate influences nitrogen balance, dry matter digestibility, and carcass traits in growing-finishing pigs*. J Journal of animal science, 1997. **75**(5): p. 1319-1323.
- [21]. Moonsie-Shageer, S. and D. Mowat, *Effect of level of supplemental chromium on performance, serum constituents, and immune status of stressed feeder calves*. J Journal of Animal Science, 1993. **71**(1): p. 232-238.
- [22]. Wang, M., et al., *Efficacy of Cr (III) supplementation on growth, carcass composition, blood metabolites, and endocrine parameters in finishing pigs*. J Asian-Australasian Journal of Animal Sciences, 2009. **22**(10): p. 1414-1419.
- [23]. Soltan, M., et al., *Effect of dietary chromium supplementation on growth performance, rumen fermentation characteristics and some blood serum units of fattening dairy calves under heat stress*. 2012. **11**(9): p. 751-756.
- [24]. Al-Saiady, M., et al., *Effect of chelated chromium supplementation on lactation performance and blood parameters of Holstein cows under heat stress*. J Animal Feed Science Technology, 2004. **117**(3-4): p. 223-233.
- [25]. Krajničáková, M., et al., *Selected clinico-biochemical parameters in the puerperal period of goats*. J Bull. Vet. Inst. Pulawy, 2003. **47**: p. 177-182.
- [26]. Esa, R., A. , *Effect of vitamin E and selenium supplementation on reproductive efficiency and immune response of Baladi goats under Southern Sinai*. . Ph.D., thesis, Women's Collage for Arts, Science and Education, Ain Shams Univ., Cairo, Egypt., 2011.
- [27]. Soliman, E., A. El-Moty, and A. Kassab, *Combined effect of vitamin E and selenium on some productive and physiological characteristics of ewes and their lambs during suckling period*. J Egyptian Journal of Sheep Goat Sciences, 2012. **7**(2): p. 31-42.

- [28]. Kumar, N., et al., *Selenium supplementation influences growth performance, antioxidant status and immune response in lambs*. J Animal Feed Science Technology 2009. **153**(1-2): p. 77-87.
- [29]. El-Shahat, K. and U. Abdel Monem, *Effects of dietary supplementation with vitamin E and/or selenium on metabolic and reproductive performance of Egyptian Baladi ewes under subtropical conditions*. J World Applied Sciences Journal, 2011. **12**(9): p. 1492-1499.
- [30]. Mahmoud, G.B., S.M. Abdel-Raheem, and H.A. Hussein, *Effect of combination of vitamin E and selenium injections on reproductive performance and blood parameters of Ossimi rams*. J Small Ruminant Research, 2013. **113**(1): p. 103-108.
- [31]. Hamam, A. and H. Abou-Zeina, *Effect of vitamin E and selenium supplements on the antioxidant markers and immune status in sheep*. J. Biol. Sci, 2007. **7**(6): p. 870-878.
- [32]. Chang, X. and D. Mowat, *Supplemental chromium for stressed and growing feeder calves*. J Journal of Animal Science, 1992. **70**(2): p. 559-565.
- [33]. Yang, W., et al., *Effects of chromium supplementation on early lactation performance of Holstein cows*. J Canadian Journal of Animal Science, 1996. **76**(2): p. 221-230.
- [34]. Pechova, A. and L. Pavlata, *Chromium as an essential nutrient: a review*. J Veterinarni Medicina-Praha-, 2007. **52**(1): p. 1.
- [35]. Nikkhah, A., et al., *Chromium improves production and alters metabolism of early lactation cows in summer*. J Journal of animal physiology animal nutrition 2011. **95**(1): p. 81-89.
- [36]. Malik, M., et al., *The effect of supplemental dietary chromium on blood glucose, body weight and liver enzymes of rabbits*. J Journal of Medicinal Plants Research, 2011. **5**(16): p. 3940-3945.
- [37]. Tahmasbi, M., et al., *Effects of selenium and vitamin E and night or day feeding on performance of Holstein dairy cows during hot weather*. Journal of Cell Animal Biology, 2012. **6**(3): p. 33-40.
- [38]. Okab, A., et al., *Seasonal changes in plasma thyroid hormones, total lipids, cholesterol and serum transaminases during pregnancy and at parturition in Barki and Rahmani ewes*. J Indian Journal of Animal Sciences, 1993.
- [39]. El-Masry, K., M. Nessim, and A. Gad, *Determination of heat tolerance coefficient in crossbred and Baladi pregnant cows under Egyptian environmental conditions*. J. Rad. Res. Applie. Sci, 2010. **3**(4): p. 1399-1409.
- [40]. Sharma, A. and K. Nalini, *Effect of extreme hot climate on liver and serum enzymes in Marwari goats*. J Indian Journal of Animal Sciences, 2011. **81**(3): p. 293-295.
- [41]. Marai, I., et al., *Effects of Egyptian subtropical summer conditions and the heat-stress alleviation technique of water spray and a diaphoretic on the growth and physiological functions of Friesian calves*. J Journal of Arid Environments, 1995. **30**(2): p. 219-225.
- [42]. Paul, T.K., S. Haldar, and T.K. Ghosh, *Growth performance and nutrient utilization in black Bengal bucks (Capra hircus) supplemented with graded doses of chromium as chromium chloride hexahydrate*. J Journal of Veterinary Science, 2005. **6**(1): p. 33-40.
- [43]. Yazaki, Y., et al., *A pilot study of chromium picolinate for weight loss*. J The Journal of Alternative Complementary Medicine, 2010. **16**(3): p. 291-299.
- [44]. Al-Bandr, L.K., D.K. Ibrahim, and E.H. Al-Mashhadani, *Effect of supplementing different sources of Chromium to diet on some physiological traits of Broiler Chickens*. J Egyptian Poultry Science, 2010. **30**(2): p. 397-413.
- [45]. Sashidhar, G. and T. Prasad, *Influence of selenite and selenomethionine administration on serum transaminases and haematology of goats*. J Indian Journal of Animal Nutrition, 1993. **10**(1): p. 1-6.
- [46]. Singh, R., S. Randhawa, and K. Dhillon, *Changes in blood biochemical and enzyme profile in experimental chronic selenosis in buffalo calves (Bubalus bubalis)*. J Indian Journal of Animal Sciences, 2002. **72**(3): p. 230-232.
- [47]. Saleh, E.M., *Studies on vitamin E deficiency in lambs*. . MSc., thesis, Fac. Vet. Med., Cairo Univ., Cairo, Egypt., 2000.
- [48]. Garg, D.P., et al., *Role of vitamin E in mitigating methomyl induced acute toxicity in blood of male Wistar rats*. J Drug chemical toxicology, 2008. **31**(4): p. 487-499.
- [49]. Ibrahim, W., et al., *Effects of dietary carnosine and vitamin E on antioxidant and oxidative status of rats*. J International journal for vitamin nutrition research, 2008. **78**(45): p. 230-237.
- [50]. Korieim, K., M.S. Arbid, and N. El-Gendy, *The protective effect of some antioxidants against the toxic effect of the alcoholic extract of faba beans on albino rats*. J Rev Latinoamer Quím, 2009. **37**: p. 181-93.
- [51]. Alameen, A.O. and A.M. Abdelatif, *Metabolic and endocrine responses of crossbred dairy cows in relation to pregnancy and season under tropical conditions*. J American-Eurasian Journal of Agricultural Environmental Science, 2012. **12**(8): p. 1065-1074.
- [52]. Nazifi, S., M. Saeb, and S. Ghavami, *Serum lipid profile in iranian fat- tailed sheep in late pregnancy, at parturition and during the post- parturition period*. J Journal of Veterinary Medicine Series A, 2002. **49**(1): p. 9-12.
- [53]. Marai, I., et al., *Reproductive performance traits as affected by heat stress and its alleviation in sheep*. J Tropical Subtropical Agroecosystems, 2008. **8**(3): p. 209-234.
- [54]. Watson, T., et al., *Effects of pregnancy and lactation on plasma lipid and lipoprotein concentrations, lipoprotein composition and post-heparin lipase activities in Shetland pony mares*. J Reproduction, 1993. **97**(2): p. 563-568.
- [55]. Ocak, S. and G. Okan, *Physiological responses and some blood parameters of bucks under Mediterranean climate conditions*. J Anadolu Tarım Bilimleri Dergisi, 2010. **25**(2): p. 113-119.
- [56]. Pandey, N., et al., *Ambient stress associated variations in metabolic responses of Marwari goat of arid tracts in India*. J Journal of Stress Physiology Biochemistry, 2012. **8**(3).
- [57]. Gudev, D., et al., *Effect of heat-stress on some physiological and biochemical parameters in buffaloes*. J Italian Journal of Animal Science, 2007. **6**(sup2): p. 1325-1328.
- [58]. Bunting, L., et al., *Influence of chromium picolinate on glucose usage and metabolic criteria in growing Holstein calves*. J Journal of Animal Science, 1994. **72**(6): p. 1591-1599.
- [59]. Depew, C., et al., *Performance and metabolic responses of young dairy calves fed diets supplemented with chromium tripicolinate*. J Journal of Dairy Science, 1998. **81**(11): p. 2916-2923.
- [60]. Brzoska, F. and B. Brzoska, *Effect of dietary selenium on milk yield of cows and chemical composition of milk and blood*. J Annals of Animal Science, 2004. **4**(1).
- [61]. Kalmath, G.P., M.N. Swamy, and S. Yathiraj, *Effect of summer stress and supplementation of vitamin E and selenium on serum lipid profile in Hallikar cattle*. J Int. J. Sci. Res, 2013. **4**: p. 95-97.
- [62]. Kumar, M., R. Jindal, and S. Nayyar, *Physiological and biochemical profile of summer stressed goats*. J Indian Veterinary Journal, 2012. **89**(8): p. 38.
- [63]. Sejjan, V., S. Indu, and S. Naqvi, *Impact of short term exposure to different environmental temperature on the blood biochemical and endocrine responses of Malpura ewes under semi-arid tropical environment*. J Indian J. Anim. Sci, 2013. **83**(11): p. 1155-1160.
- [64]. Das, K., et al., *Effect of heat stress alleviation on plasma protein, metabolites and lipid profile in lactating Nili-Ravi buffaloes under tropical climate*. J Indian J. Anim. Sci, 2013. **83**(5): p. 86-89.

- [65]. Harper, H.A., Rowell, V.W. and Mayes, P.A., *Review of physiological chemistry*. 17th ed. PP. 511. Middle East Edition, Lange medical publications, Beirut, Lebanon., 1971.
- [66]. Dixon, R., R. Thomas, and J. Holmes, *Interactions between heat stress and nutrition in sheep fed roughage diets*. J The Journal of Agricultural Science, 1999. **132**(3): p. 351-359.
- [67]. Marai, I. and A. Habeeb, *Adaptability of Bos taurus cattle under hot arid conditions*. J Annals of Arid Zone, 1998. **37**(3): p. 253-281.
- [68]. Habeeb, A.A.M., Marai, I. F. M. and Kamal, T. H., *Heat stress*. . In: C. Philips and D. Piggens, editors, *Farm animals and the environment*. CAB International, Wallingford, UK. , 1992: p. 27–47.
- [69]. Shwartz, G., et al., *Effects of a supplemental yeast culture on heat-stressed lactating Holstein cows*. J Journal of dairy science, 2009. **92**(3): p. 935-942.
- [70]. Montmurro, N., C. Pacelli, and A. Borghese, *Metabolic profiles in buffalo heifers bred in two farms with different feeding and climatic conditions*. J Egyptian J. Anim. Prod, 1995. **32**(1): p. 1-12.
- [71]. Yanchev, I., et al., *Effect of Cr picolinate and Zn supplementation on plasma cortisol and some metabolite levels in Charolais hoggets during acclimatization*. J Archiva Zootechnica, 2007. **10**: p. 78-84.
- [72]. Ozawa, M., et al., *Alterations in follicular dynamics and steroidogenic abilities induced by heat stress during follicular recruitment in goats*. J Reproduction, 2005. **129**(5): p. 621-630.
- [73]. Hein, K. and R. Allrich, *Influence of exogenous adrenocorticotrophic hormone on estrous behavior in cattle*. J Journal of animal science, 1992. **70**(1): p. 243-247.
- [74]. Roth, Z., *Heat stress, the follicle, and its enclosed oocyte: mechanisms and potential strategies to improve fertility in dairy cows*. J Reproduction in Domestic Animals, 2008. **43**: p. 238-244.
- [75]. Balicka-Ramis, A., et al., *Effects of selenium administration on blood serum Se content and on selected reproductive characteristics of sheep*. J Archives Animal Breeding, 2006. **49**(2): p. 176-180.
- [76]. Gilad, E., et al., *Effect of heat stress on tonic and GnRH-induced gonadotrophin secretion in relation to concentration of oestradiol in plasma of cyclic cows*. J Reproduction, 1993. **99**(2): p. 315-321.
- [77]. Soltan, M., et al., *Effect of dietary chromium supplementation on growth performance, rumen fermentation characteristics and some blood serum units of fattening dairy calves under heat stress*. J Pakistan Journal of Nutrition, 2012. **11**(9): p. 751-756.
- [78]. Tuormaa, T.E., *Chromium, selenium and copper and other trace minerals in health and reproduction*. J Journal of orthomolecular medicine, 2000. **15**(3): p. 145-156.
- [79]. Pradini, W., S. Rahayu, and M. Djati, *Level of estrogen and cervical mucus pH as indicator of estrus after calving towards the provision of selenium vitamin ETM on dairy cow Friesien Holstein (FH)*. J International Journal of ChemTech Research, 2015. **7**(1): p. 190-195.
- [80]. Nayyar, S. and R. Jindal, *Essentiality of antioxidant vitamins for ruminants in relation to stress and reproduction*. J Iranian Journal of Veterinary Research, 2010. **11**(1): p. 1-9.

Dr. S. S. Emara, et. al. "Physiological and blood biochemical responses in Baladi Female Goats treated with Chromium and Selenium-E under hot summer conditions." *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, 14(2), 2021, pp. 43-56.