# Phytosocial Diversity and Distribution of Herbaceous Species in Dryland Ecosystem of Kebbi, North-western Nigeria

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## Abstract:

**Background:** Phytosociology describe species population dynamics and their relationship to other species. This study aimed at determine phytosocial diversity and distribution of herbaceous species. Materials and Methods: Herbaceous species were determined in randomly selected 400 quadrats  $(1m \times 1m)$  by identifying, analyzing frequency and evaluation species diversity using various indices. **Results:** A total of 4372 herbaceous species belonging to 44 genera were distributed in 18 families. Fabaceae recorded the highest species occurrence (13.3%), followed by Poaceae and Amaranthaceae each with 11.1%, while the rare species were found under Asteraceae, Cucurbiataceae, Rubiaceae, Cleomarceae, Cyperaceae, and family Phyllanthaceae, each with 2.22% relative frequency. Site A recorded the highest species 34.90%, followed by Site C, D and B with 25.07%, 21.55% and 18.32% respectively (P<0.05). Teprosia pedicelata had the highest species distribution with 8.71%, followed by Mitracarpus scabrunzuce, and Leucus martinicensis, with 5.38% and 3.68% respectively. Eragrotis tremula, Phyllanthus amarus and Sesbania sesban were found rare only in Site D(0.37%), A(0.09%) and C(0.34%) respectively. However, Citrilus colosynthius found in Site A, B and D shows rare distribution of 0.50%. Approximately 70.45% of herbaceous species shows aggregate distribution and 29.55% random distribution pattern. Diversity indices varies where, Simpson and Shannon index ranges from 0.941 in Site D to 0.961 in Site C and 2.941 in Site D to 3.335 in Site C respectively. Conclusion: Herbaceous species formed aggregate distribution than random distribution pattern. Environmental and human activities influence species diversity and distribution. Rare species should be conserve for future purposes by protecting their habitats.

Key words: Phytosocial, Herbaceous, Distribution, Species, Diversity, Composition.

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### I. Introduction

Phytosociology is the quantitative study of vegetation, aimed to predict its pattern of distribution<sup>1</sup>. Phytosociology refers to the characteristics, classification, relationship and distribution of plant communities of a particular environment<sup>2</sup>. It aids in forest and grassland management planning, environmental impact studies, restoration and reclamation of degraded areas, and may also indicate the potential species richness and abundance<sup>3</sup>. It is used to describe the population dynamics of species and their relationship to other species in the same community<sup>4, 2</sup>. Phytosociological studies are essential for protecting the natural plant communities and biodiversity as well as understanding the changes experienced in the past and future. Analysis of a plant community, it species composition and phytosociological interaction of species<sup>5</sup>. Anthropogenic activities like overexploitation, pollution, habitat destruction and degradation by physical and chemical means causing significant effect on the pattern of phytosocial distribution as well as biodiversity loss<sup>6</sup>. Conversion of local vegetation for other human activities is a major threat to biodiversity <sup>7, 8, 9, 10</sup>.

Herbaceous vegetation fluctuates in a cyclic manner from one season to another in a successional way and modified to prevailing factors<sup>11</sup>. These fluctuations continuous in space and time due to a multitude of factors like overgrazing, fire, soil nutrients, rainfall and human activities which differs in intensity and duration<sup>12</sup>. Plant species behave differently in natural community to the varying environmental conditions<sup>13, 14, <sup>15</sup>, either abiotic<sup>16, 17</sup> or biotic factors<sup>18</sup>, and consequently result in the evolution of diverse communities. Herbaceous species constitute up to 60% of the plants species diversity in terrestrial ecosystem<sup>19</sup>. Naturally, they serve as habitats for a wide array of animals, basis for complex food webs<sup>20, 21, 22, 23</sup>, and are involved in protecting topsoil, maintain biodiversity, improving water penetration into soils and soil holding capacity<sup>24, 25, <sup>26</sup>, mitigation of CO<sub>2</sub>, livestock feeds, runoff and wind reduction, and above-ground carbon storage<sup>27, 28, 29, 30</sup>. Despite these ecological importance of herbaceous layer, they are poorly studied and are usually not included in</sup></sup> most floristic studies <sup>31, 32</sup>. Herbaceous species interacts with different species that share similar environmental requirements to forms different vegetation types and may also respond to various environmental changes <sup>33</sup>.

Biodiversity refers to the number of different species in a particular area (species richness) weighted by abundance of individuals species. Barnes *et al.* <sup>34</sup> defined biodiversity as the kinds and numbers of organisms and their patterns of distribution in an ecosystem. However, species diversity can be measure by species evenness i.e. abundance of each species represented in an area <sup>35</sup>, and species richness; the biological measure of alpha ( $\alpha$ ) diversity, usually expressed as the number of species per unit sample <sup>36</sup>. Therefore, diversity is affected by species richness and evenness <sup>37</sup>. Diversity is an important structural attributes of a natural community which are related to functional properties of vegetation like productivity, niche structure, competition, stability and integration of the community <sup>38</sup>. Species diversity is minimum when individuals are of one species and maximum when individuals belong to a larger number of different species <sup>39</sup>. The assemblage of plants found at a given locality can be interpreted as the products of filtering the effects of climatic conditions, edaphic factors, biotic interactions and type of disturbance <sup>40</sup>. The development and deterioration of plant species alters the pattern of species distribution in community<sup>11</sup>. Increased in biodiversity has been found to increase primary productivity <sup>41</sup>, changes plant allocation pattern <sup>42</sup>, and reduce invisibility by unsown species that changing herbage composition <sup>42, 43</sup>. Biodiversity and distribution provide basic information for decision-making in management and conservation efforts and also provide information on the floristic composition and structures of vegetation <sup>44</sup>.

Shannon-Weiner Index and Simpson Indices are the most common indices for studying species diversity <sup>45, 46</sup>, while species richness and evenness are the components of biodiversity <sup>47 48</sup>. Simpson index is used to assess the dominance, while Shannon-Wiener index help to determine species evenness and richness, but does not provide information on rare species, which are also important <sup>49</sup>. This indicates that diversity cannot be determined by just one indices <sup>50, 51</sup>. Diversity indices are used as indicators of the community's composition and the effects of environmental changes on species <sup>37</sup>. Scientific studies on biodiversity is necessity in preserving, restoring and proper management of the existing plant species for their ecological values. The structural property of a community is the quantitative relationship between the species growing around. Assessments of plant diversity and vegetation composition of fragmented habitats to habitat fragmentation. This research was aimed at studying the current status of phytosocial diversity and distribution of herbaceous species in dryland ecosystem.

#### **Study Location**

## II. Materials and Methods

Kebbi state is located in North-western Nigeria, between latitude 12.45<sup>o</sup> N and longitude 4.2<sup>o</sup> E. It covers a geographical land area of 36,800 km<sup>2</sup>, borders with nation of Niger Republic to the west and Benin Republic to the South-east and locally bordered Nigerian states; Niger, Sokoto, Zamfara to the South, North and East respectively. The state marked by a single rainy season last from May to October with mean annual rainfall of about 720mm and long dry season last for the remaining period of the year <sup>52</sup>. The mean temperature range is 26<sup>o</sup>C during harmattan season (November to February) and 38<sup>o</sup>C-40<sup>o</sup>C during the month of April to June <sup>53</sup>. Its vegetation is Northern Guinea Savanna in the South and South-East and Sudan Savanna in the North, covers with short grasses and small trees. Although, the state face desertification but many areas are still being altered by cultivation, grazing, cutting of fuel woods, excavation of soil, bush fire and so on. The study was conducted in four Sites including Aleiro located between longitude 4.4388<sup>o</sup> E and Latitude 12.3562<sup>o</sup> N, Kalgo 4.2000<sup>o</sup> E and Latitude 12.3342<sup>o</sup> N, Argungu 4.5367<sup>o</sup> E and Latitude 12.7495<sup>o</sup> N and Bunza 4.0108<sup>o</sup> E and Latitude 12.0916<sup>o</sup> N, all the study Sites was protected from anthropogenic activities during the period of research, but all the study Sites experienced various anthropogenic activities before the research was commence.

#### Study Design

Phyto-social diversity and distribution of herbaceous species were conducted during the month of June to October, 2020. Completely Randomize Design (CRD) was adopted for monthly sampling due homogeneity of the experimental Sites, twenty 20 ( $1m^2 x 1m^2$  quadrats) on each sampling date per plot were randomly chosen using an online random number table for generating 400 ( $1m^2 x 1m^2$  quadrats) throughout the study periods and plants species occur at the 20 x 20cm central square quadrats were identified <sup>54</sup>.

#### III. Methodology

Twelve plots (30m x 30m) were established and marked out with iron rods (2 meter above the ground), for monthly sampling from June to October, 2020. The plots were divided into quarters and each quarter were then divided into grids of 1m x 1m quadrats to give a total of 225 quadrats <sup>54</sup>. Phytosocial compositions of herbaceous species were identified, collected, pressed, dried and stitched on standard herbarium sheets of  $28 \times$ 

42 cm according to Jain and Rao <sup>55</sup>. Plant species occur in the central 20cm x 20cm randomly selected quadrat were identified *in-situ* based on their morphological, structural and floral characteristics with the aid of West African Weeds <sup>56</sup> and Flora of West Tropical Africa <sup>57, 58</sup>. The *in-situ* identified plants were transported to Herbarium, Department of Biology, Federal University Birnin Kebbi (FUBK), Nigeria, for authentication of species.

#### **Statistical Analysis**

Analysis of variance (One-way ANOVA) were used for analyzing species composition and distribution using MINITAB (Version 18) at 95% confidence interval. Diversity Indices were calculated using the software Community Ecology Parameter Calculator (ComEcoPaC) Version 2.0. Various species diversity indices were determined through Shannon wiener index (H) and community dominance index (CDI) based on the following formula;  $H = - [\sum Pi InPi]$ , where H = diversity index, Pi = proportion of each species in the sample, InPi = natural logarithm of this proportion, while  $CDI = \frac{Y1+Y2}{Y}$ , where  $Y_1 =$  most dominant species,  $Y_2$ = second most dominant species and Y= total number of species recorded.

#### IV. Results

During the study period, 44 genera of herbaceous plants species were identified and distributed over 18 families. Fabaceae recorded the highest species occurrence with (13.3%) relative frequency, followed by Poaceae and Amaranthaceae each with 11.1%, while family Malvaceae recorded 8.89%, Euphobiaceae, Solanaceae, and Lamiaceae each had 6.67% relative frequency, and the rare species were recorded under family Asteraceae, Cucurbiataceae, Rubiaceae, Cleomaceae, Cyperaceae, and Phyllanthaceae, each with 2.22% relative frequency. Table 1 shows the distribution and abundance of 4372 herbaceous species in four study Sites. Sites A recorded the highest species composition of 34.90% relative frequency, followed by Site C and D with 25.07% and 21.71% respectively, while least species composition were recorded under Site B with 18.32% enumerated species. The distribution and abundance of species between the Sites were statistically significance (p<0.05) at 95% confidence interval (Table 2).

Table 1: Relative Frequency of Herbaceous Species Composition and Distribution Identified in the Four Study Sites, 2020

		5	Sites and Species RF (%)			Total RF	
Family	Species Name	А	В	С	D	(%)	
Acanthaceae	Hygrophila auriculata (L.) Schum	0.71	0.21	0.43	0.89	2.24	
	Blepharis maderaspatensis L.	0.43	0.00	0.85	0.41	1.69	
Amaranthaceae	Amaranthus viridis L.	0.66	0.85	0.94	0.00	2.45	
	Celosia trigyna L.	0.39	0.71	0.00	1.26	2.36	
	Achyrathes aspera L.	0.00	0.48	1.17	0.00	1.65	
	Gomphrena celosiodes Mart	0.00	1.08	1.10	0.00	2.17	
Asteraceae	Acanthospermun hispidum Dc	0.89	0.98	0.00	1.58	3.45	
Cleomaceae	Cleome viscosa L.	0.85	0.00	0.85	0.00	1.69	
Ceasalpiniaceae	Cassia occidentalis L.	2.01	1.17	0.16	0.00	3.34	
	Cassia tora L.	0.25	0.11	1.21	0.80	2.38	
Convolvulaceae	Evolulus alsinoides L.	0.80	0.00	0.07	0.94	1.81	
	Ipomoea muricata L.	1.40	0.94	0.43	0.00	2.77	
Cucurbiataceae	Citrilus colosynthus L.	0.16	0.32	0.00	0.02	0.50	
Commelinaceae	Cyanotis lanata L.	0.98	0.00	0.82	0.00	1.81	
	Commelina errecta L.	0.94	0.66	0.75	0.07	2.42	
Cyperaceae	Cyperus rotundus L.	1.78	0.00	0.00	1.30	3.09	
Euphorbiaceae	Euphorbia hirta, L.	0.07	0.00	1.33	1.01	2.40	
	Phyllanthus pentendrus Sch and thon	1.17	1.26	0.00	0.00	2.42	
	Chrozophora brocchiana Schweinf.	0.75	0.00	0.46	0.00	1.21	
Fabaceae	Teprosia pedicelata Baker	4.14	1.42	0.94	2.22	8.71	

	Crotolaria mucronata Desv	0.46	0.00	0.59	0.00	1.05
	Alysicarpus vaginalis L.	1.08	1.05	0.00	1.26	3.39
	Cassia mimosoides L.	0.82	0.00	0.50	0.00	1.33
	Tephrosia linearis (wild) Pers.	1.58	0.00	0.30	1.90	3.77
	Sesbania sesban L.	0.00	0.00	0.34	0.00	0.34
Lamiaceae	Hyptis spicigera L.	1.24	0.00	0.94	0.00	2.17
	Ocimum basilicum L.	0.41	0.00	0.00	0.59	1.01
	Leucus martinicensis (Jacq) R.	1.51	0.94	1.24	0.00	3.68
Malvaceae	Waltheria indica L.	0.94	0.32	0.89	0.62	2.77
	Urena lobata L.	0.46	0.00	1.78	1.05	3.29
	Sida acuta L.	1.40	0.00	0.64	0.00	2.04
	Corchorus olitorius L.	0.00	0.30	0.98	0.73	2.01
Phyllanthaceae	Phyllanthus amarus	0.09	0.00	0.00	0.00	0.09
Poaceae	Cynodon dactylon (L.) Pers.	0.00	1.26	0.25	0.78	2.29
	Eleusin indica (L.) Gaertn	1.74	0.00	1.30	0.00	3.04
	Pennisetum pedicellatum Trim	0.66	1.12	0.98	0.00	2.77
	Eragrotis tremula Hochst	0.00	0.00	0.00	0.37	0.37
	Digitaria debelis Haller	0.78	0.00	1.10	0.57	2.45
Portulaceae	Portulaca olerceae L.	0.78	0.50	0.00	0.00	1.28
	Securidaca longepedunculata Fres	0.00	0.00	0.00	0.30	0.30
Rubiaceae	Mitracarpus scabrunzuce	1.94	1.58	0.53	1.33	5.38
Solanaceae	Physalis angulate L.	0.00	0.00	0.00	1.72	1.72
	Schweinkia americana L.	0.00	0.00	0.48	0.00	0.48
	Solanum nigrrum L.	0.64	1.08	0.71	0.00	2.42
		34.90	18.32	25.07	21.71	100.00

Keys: RF= Relative Frequency, Site; A= Aliero, B= Kalgo, C= Argungu and D= Bunza

Among the species *Teprosia pedicelata* had the highest number of species with 8.71% relative frequency (Table 1), followed by *Mitracarpus scabrunzuce*, and *Leucus martinicensis*, with relative frequency of 5.38% and 3.68% respectively. While *Eragrotis tremula* and *Securidaca longepedunculata* was found rare only in Site D with 0.37% and 0.30% respectively, *Phyllanthus amarus*, and *Schweinkia Americana* and *Sesbania sesban* were only recorded in Site A(0.09%), and D(0.48%) and (0.34%) respectively. However, *Citrilus colosyntnthus* found in Site A, B and D but shows lowest species composition of 0.50% relative frequency. Regards to phytosocial distributions, the results shows that most annual plant species shows aggregate distribution pattern with approximately 70.45%, few species revealed that 29.55% were randomly distributed across the study area.

	<b>Table 2</b> : O	ne-Way Analysi	s of Variance (ANOV	A)	
Source of Variation	AdjSS	Df	AdjMS	F	P-Value
Sites	8520.297	3	2840.099	4.150251	0.007226
Species	114965.7	168	684.3198		
Total	123486	171			

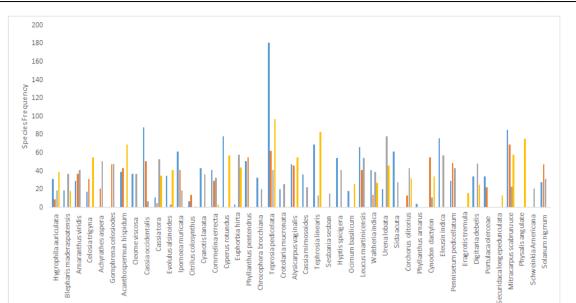


Fig. 1: Species Composition and Distribution in Four Study Sites

C D

Table 3 shows that, the diversity indices varies between the study Sites, Simpson and Shannon diversity index varied from 0.941 in Site D to 0.961 in Site C and 2.941 in Site D to 3.335 in Site C respectively, while species evenness found to be higher in Site C (E= 0.878) and lower in Site A (E= 0.720). The species dominance was found maximum in Site D (D= 0.059) and minimum in Site C (D= 0.039). The results indicates that, the herbaceous communities of the study Sites experience less environmental stress and the species shows similar distribution and abundance since Simpson diversity index is closer to 1. CDI value ranges from minimum of 0.164 in Site B to maximum of 0.189 in Site D, Menhinick diversity index value were found between 0.747 in site D and 0.967 in site C and Margalef Index ranges from 3.141 in site B to 4.638 in site A respectively. Equitability index shows similar values but was found minimum (0.938) in site D and maximum (0.962) in site C. Berger-parker and Fisher alpha index recorded the highest value of 0.1186 in site A, and lowest value (0.086) value in site C and 6.386 in site A and lowest value (4.249) in site D.

Indices	Sites					
	А	В	С	D		
Dominance	0.044	0.056	0.039	0.059		
Simpson	0.956	0.944	0.961	0.941		
Shannon	3.320	2.954	3.335	2.941		
Evenness	0.790	0.872	0.878	0.834		
Brillouin	3.262	2.887	3.263	2.883		
CDI	0.176	0.164	0.124	0.189		
Menhinick	0.896	0.777	0.967	0.747		
Margalef	4.638	3.141	4.429	3.209		
Equitability	0.939	0.956	0.962	0.938		
Berger-parker	0.119	0.086	0.089	0.102		
Fisher alpha	6.386	4.182	6.172	4.249		

Note: CDI= Community dominance Index

# V. Discussion

In this study, the results showed that phytosocial diversity and distribution differed among the species as well as between the study sites (A, B, C and D), which may be influenced by human activities and soil condition. Composition and distribution of species was found to be higher (34.90%) in Site A compared to the

others Sites. High composition and distribution in Site A, indicates the availability of soil nutrients that favors the composition and distribution of herbaceous species and also indicates its primary productivity. The lower species composition in Site B (18.32%) may be due human activities, low soil nutrients and production of less number of seeds for germination since availability of seeds favors the composition and distribution of species. Larger species composition are more able to reach multiple patches of habitat within a landscape to find the resources they need <sup>59</sup>. Plants species composition is associated with soil nutrients and other environmental resources that are needed to species richness<sup>60, 61</sup>. *Teprosia pedicelata* with highest (8.71%) number of species might indicates it adaptability of the environmental condition and also higher seeds production during the growing season, while Eragrotis tremula, Securidaca longepedunculata, Phyllanthus amarus, Schweinkia americana and Sesbania sesban with 0.37%, 0.30%, 0.09%, 0.48% and 0.34% respectively, are found rare during the study area and occurs only in a single sites, which may be due the influence of environmental factor and limited seeds dispersal. This study is in line with the findings of Casas and Ninot <sup>33</sup> and Alados et al. <sup>40</sup> that said the assemblage of plants found at any given locality is a product of filtering the effects of edaphic factors and disturbance type. Citrilus colosynthtus found in Site A, B and D shows lower (0.50%) species composition, which might indicates high environmental sensitivity and or less seed production and limited seeds dispersal. Every plant species has a tolerance limit that expand or narrow its distribution and may form mutual relationship with different species that share similar environmental requirements <sup>4, 62</sup>. Anthropogenic disturbances critically affect the biodiversity and the structural characteristics of the community <sup>63, 64</sup>. Nakahama *et al.* <sup>65</sup> reported that, anthropogenic activities such as urbanization, industrialization, over-grazing, salinization, solid wastes, military activities, over-cutting of woody plants, road construction, and establishment of new settlements are the main drivers of change, transformation, and loss of natural habitats; decline in floristic composition and dramatic changes in vegetation structure. In this study, the results shows that most of the herbaceous species shows aggregate distribution pattern with approximately 70.45%, few species revealed that 29.55% were randomly distributed across the study area, this may be due to aggregates seeds dispersal or environmental resources. The results are in line with Das et al. 66 that said aggregated distribution indicated habitat preference, while random distribution indicates the environment in which plant species grow is homogeneous and has many factors acting on the population <sup>36</sup>.

Simpson and Shannon diversity index varied from 0.941 in Site D to 0.961 in Site C and 2.941 in Site D to 3.335 in Site C respectively, while species evenness found to be higher in Site C (E=0.878) and lower in Site A (E= 0.720). The differences in the phytosociological parameters may be attributed to different biotic and/or abiotic factors other than soil and elevation. Menhinick diversity index value were found between 0.747 in site D and 0.967 in site C and Margalef Index ranges from 3.141 in site B to 4.638 in site A respectively. The low diversity recorded in site D(H= 2.941) and B(H= 2.954), may be attributed to lesser number of species, the community is dominated by few individuals or may be due to environmental degradation as a result of anthropogenic pressures. Simpson diversity index is always higher where the community is dominated by less number of species and when the dominance is shared by large number of species <sup>61</sup>. This indicate that Shannon diversity index of this research is similar to the value of the research conducted in Savanna ecosystem by Salisu and Rabiu <sup>61</sup>, where he found 'H' ranges from 2.63 to 3.10. Hooper *et al.* <sup>39</sup> reported that species diversity is minimum when the individuals are of one species and maximum when individuals belongs to a large number of different species. According to Bhandari *et al.*<sup>67</sup>, every species in a community plays a specific role and there is a definite quantitative relationship between abundant and rare species. Wilsey and Stirling 68, reported that richness and evenness can be negatively related across the plant communities, and evenness can account for more variation in Shannon's diversity index (H) than richness, which suggests that relationships among the diversity components can be complex. Dash <sup>69</sup>, reported that matured and stable communities have high diversity value (0.6 to 0.9), while the communities under stress conditions, exhibiting low diversity, usually show close to zero value. Margalef index uses species richness to compare variation of a community and reflecting sensitivity to sample size, while Menhinick index used to estimate species richness but is independent on the sample size <sup>70</sup>. The Maximum value 0.119 and minimum value 0.086 of Berger-Parker index recorded in site A and B respectively in this study, in line with Shannon Wiener Index of this study. Javaid and Asho  $^{70}$ , reported that the higher the value of Berger-Parker index, the higher the diversity and lower the dominance.

# VI. Conclusion

The result of this research shows that, phytosocial diversity and distribution of herbaceous species are favored by larger seeds production, limited seeds dispersal and availability of soil nutrients, while anthropogenic activities and environmental pressure distract phytosocial diversity and distribution. The research revealed that, dryland area is not suitable for the growth and development of *Eragrotis tremula*, *Securidaca longepedunculata*, *Phyllanthus amarus*, *Schweinkia americana* and *Sesbania sesban*, are found rare and occurs only in a single sites. The diversity indices of this study shows that the study area is more or less stable for most herbaceous community, although some study sites (B and D) have low values of diversity indices due to environmental

pressure and anthropogenic activities. The study on the influence of edaphic factors on physocial diversity and distribution should be conducted.

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#### References

- [1]. Ilorkar VM, Khatri PK. Phytosociological Study of Navegaon National Park, Maharashtra. India Forester, 2003; 129(3): 377-387.
- [2]. Rao DS, Prayaga PM, Aniel OK. Plant Biodiversity and Phytosociological Studies on Tree Species diversity of Khammam District, Telangana State, India, *Journal of Pharmaceutical Science and Research*, 2015; 7(8): 518-522
- [3]. Santos WS, Souza MP, Nobrega GFQ, Medeiros FS, Alves AR, Hokanda, AC. Floristic-phytosociological characterization of the woody component In a caatinga fragment in the municipality of Upanema-RN. *Native Sinop*, 2017; 5(2): 85-91. DOI: 10.5935 / 2318-7670.v05n02a02
- [4]. Mishra NK, Singh R, Ojha S, Supreeti S. Phytosociological perspectives of representative herbaceous genera of common occurrence belonging to family asteraceae in grassland ecosystem of Anpara Region in district Sonebhadra (U.P.). Indian Journal of Life Science, 2012; 2(1):119-122.
- [5]. Mueller-Dombois D, Ellenberg H. Aims and Methods of Vegetation Ecology. John Wiley and Sons, United Stste of America. 1974; Pp: 62-65.
- [6]. Hezagy T. Optimization of resource allocation and leveling using genetic algorithms. Journal of Construction Engineering and Management, 1999; 125(3): 167-175. <u>https://doi.org/10.1061/(ASCE)07333-9364(1999)125:3(167)</u>.
- [7]. Tittensor DP, Walpole M, Hill SL, Boyce DG, Britten GL, Burgess ND, Baumung R. A mid-term analysis of progress toward international biodiversity targets. *Science*, 2014; 346(6206):241–244
- [8]. Newbold T, Hudson LN, Hill SL, Contu S, Lysenko I, Senior RA, Day J. Global effects of land use on local terrestrial biodiversity. *Nature*, 2015; 520(7545):45-52.
- [9]. Maxwell SL, Fuller RA, Brooks TM, Watson JE. Biodiversity: the ravages of guns, nets and bulldozers. Nature, 2016; 536(7615):143-145.
- [10]. Venter O, Sanderson EW, Magrach A, Allan JR, Beher J, Jones KR, Levy MA. Sixteen years of change in the global terrestrial human footprint and implications for biodiversity conservation. *Natural Community*, 2016; 7:125-158.
- [11]. Shahid M, Joshi SP. Phytosociological assessment & distribution patterns of tree species in the forests of Doon Valley, Shivalik hills of lower Himalaya, *Society for Tropical Plants Research*, 2016; 3(2): 263–271.
- [12]. Shameem SA, Soni P, Bhat GA. Comparative study of herb layer diversity in lower Dachigam National Park, Kashmir Himalaya, India. International Journal of Biodiversity and Conservation, 2010; 2(10): 308-315.
- [13]. Soininen J. Environmental and spatial control of freshwater diatoms a review. Diatom Research, 2007; 22:473-490.
- [14]. Shah JA, Pandit AK, Shah GM. Distribution, diversity and abundance of copepod zooplankton of Wular Lake, Kashmir Himalaya, *Journal of Ecology and Natural Environment*, 2013; 5(2):24-29
- [15]. Shah JA, Pandit AK. Relation between physico-chemical limnology and crustacean community in Wular lake of Kashmir Himalaya, Pakistan Journal of Biological Sciences, 2013; 16(19):976-983.
- [16]. Rindi F, Battelli C. Spatio-temporal variability of intertidal algal assemblages of the Slovenian coast (Gulf of Trieste, northern Adriatic Sea). Botany and Marine, 2005; 48:96–105.
- [17]. Charles DF, Acker FW, Hart DD, Reimer CW, Cotter PB. Largescale regional variation in diatom-water chemistry relationships: Rivers of the eastern United States. *Hydrobiologia*, 2006; 561:27–57
- [18]. Coleman MA. Small-scale spatial variability in intertidal and subtidal turfing algal assemblages and the temporal generality of these patterns. *Journal of Expert Marine Biology and Ecology*, 2002; 267:53–74.
- [19]. Mohammed AH, Jahun SF, Mohammed GA Dangana AS. Herbaceous species diversity in Kanawa forest reserve (KFR) in Gombe state, Nigeria. *American Journal of Agriculture and Forestry*, 2015; 3(4): 140-150.
- [20]. Smith BD. General Patterns of niche construction and the management of 'wild' plant and animal resources by small-scale preindustrial societies. *Philosophy and Transitional Research Sociology and Biology*, 2011; 366(1566): 836–848
- [21]. Choy SY, Prasad KMN, Wu TY Ramanan RN. A review on common vegetables and legumes as promising plant-based natural coagulants in water clarification. *International Journal of Environmental Science and Technology*, 2015; 12: 367-390.
- [22]. Andarwulan N, Faridah DN, Prabekti YS, Fadhilatunnur H, Mualim L, Aziz SA Cisneros-Zevallos L. Dietary fiber content of waterleaf (Talinum triangulare (Jacq.) wild) cultivated with organic and conventional fertilization in different seasons. American Journal of Plant Science, 2015; 6: 334-343.
- [23]. Yang Y, Ning Y, Zhu X, Li R, Ye H, Zhao L, Jin L, Zhou X. Antifungal and antiinflammatory effects of Coptidis chizoma extract against Candida albicans. *African Journal of Traditional Complementary and Alternative Medicine*, 2015; 12(4): 161-168.
  [24]. Mashwani Z, Rehman R, Qureshi R, Arshad MA, Khan MA, Ullah Z. *The diversity of grasses in the Gandgar Range, Northwest*
- [24]. Mashwani Z, Rehman R, Qureshi R, Arshad MA, Khan MA, Ullah Z. *The diversity of grasses in the Gandgar Range, Northwest Pakistan.* IC Biour-Life. 29-31 December. Center for Biodiversity and Conservation, Shah Abdul Latif University Kherpur Sindh, Pakistan, 2010; 49-57.
- [25]. Ford JD, Tilleard SE, Berrang-Ford L, Araos M, Biesbroek R, Lesnikowski AC, MacDonald GK, Hsu A, Chen C, Bizikova L. Opinion: Big data has big potential for applications to climate change adaptation. *Proceedings of the National Academy of Sciences*, 2016; 113(39): 10729–10732.
- [26]. Gilardelli F, Vergani C, Gentili R, Bonis A, Chanteloup P, Citterio S, Enrico AC. Root characteristics of herbaceous species for topsoil stabilization in restoration projects. *Land Degradation and Development*, 2017; 28(7): 2074-2085.
- [27]. Dunnett N, Qasim M. Perceived benefits to human well-being of urban gardens. *Hortcultural Technology* 2000; 10(1):40-45. http://horttech.ashspublications.org/content/10/1/40. full.pdf
- [28]. Cilliers S, Siebert S, Davoren E, Lubbe R. Social aspects of urban ecology in developing countries, with an emphasis on urban domestic gardens. *Applied Urban Ecology: A Global Framework*, 2011; 2: 123-138. http://dx.doi.org/10.1002/9781444345025. ch10.
- [29]. Davies ZG, Edmondson JL, Heinemeyer A, Leake JR, Gaston KJ. Mapping an urban ecosystem service: quantifying above-round carbon storage at a city-ide scale. *Journal of Applied Ecology*, 2011; 48:1125-1134. http://dx.doi. org/10.1111/j.1365-2664.2011.02021.
- [30]. Cameron RWF, Blanusa T, Taylor JE, Salisbury A, Halstead AJ, Henricot B, Thompson K. The domestic garden—its contribution to urban green infrastructure. *Urban Forestry and Urban Greening*, 2012; 11:129-137.

- [31]. Moro MF, Lughadha EN, Filer DL, Araujo FS Martins FR. A catalogue of the vascular plants of the Caatinga phytogeographical domain: a synthesis of floristic and phytosciological surveys. *Phytotaxa*, 2014; 160: 1-118.
- [32]. Queiroz RT, Moro MF, Loiola MIB. Evaluating the relative importance of woody versus non woody plants for alphadiversity in semi-arid ecosystem in Brazil. *Plant Ecology and Evolution*, 2015; 148(3): 361-376.
- [33]. Casas C, Ninot JM. Correlation Between species composition and soil properties in the pastures of Plana De Vic (Catalonia, Spain). Acta Botanica Barcelona, 2003; 49: 291-310
- [34]. Barnes BV, Zak DK, Denton SR, Spurr SH. Forest Ecology. John Wiley and Sons, United State O America, 1998; Pp. 773.
- [35]. Krebs CJ. Ecological Methodology. London. Chapman and Hall, 1999; Pp. 23-45.
- [36]. Ismail MI, Alawia AE. Phytosociological Analysis and Species Diversity of Herbaceous Layer in Rashad and Alabassia Localities, South Kordofan State, Sudan. Jordan Journal of Biological Sciences, 2015; 8: 151 – 157.
- [37]. Magurran A. Ecological Diversity and its Measurement. London. Chapman and Hall, 1988; Pp. 354
- [38]. Ramirez N, Dezzeo N, Chaco N. Floristic composition, plant species abundance, and soil properties of montane savannas in the Gran Sabana, Venezuela. *Flora*, 2007; 202: 316–327.
- [39]. Hooper DU, Chapin FS, Ewel JJ, Hector A, Inchausti P, Lavorel S, Lawton JH, Lodge DM, Loreau M, Naeem S, Schmid B, Setala H, Symstad AJ, Vandermeer J, Wardle DA. Effects of biodiversity on ecosystem functioning: A consensus of current knowledge, *Ecological Monographs*, 2005; 75, 3–35.
- [40]. Alados CL, ElAich A, Papanastasis VP, Ozbek H, Navarro T, Freitas H, Vrahnakis M, Larrosi D, Cabezudo B. Change in plant spatial patterns and diversity along the successional gradient of Mediterranean grazing ecosystems. *Ecological Modelling*, 2004; 180: 523–535.
- [41]. Hector A, Schmid B, Beierkuhnlein M C, Caldeira D. Plant diversity productivity experiments in European grasslands. *Science*, 1999; 286: 1123-1127
- [42]. Tilman D. Resource competition and plant traits: A response to Craine (2005). Journal Ecology, 2007; 95: 231–234.
- [43]. Kirwan L, Luscher A, Sebastia MT, Finn JA. Evenness drives consistent diversity effects in an agronomic 10 system across 28 European sites. *Journal of Ecology*, 2007; 95: 530–539
- [44]. Stachowicz JJ, Bruno JF, Duffy JE. Understanding the effects of marine biodiversity on communities and ecosystems. *Annual Review on Ecology, Evolution and Systematics*, 2007; 38:739–766.
- [45]. Souza JS, Espírito-Santo FDB, Fontes A, Oliveira-Filho AT, Botezelli L. Analysis of floristic and structural variations of the tree community of a semideciduous forest fragment on the banks of the Capivari River, Lavras - MG. *Tree Magazine of Viçosa*, 2003; 27(2):185-206.
- [46]. Gorelick R. Combining richness and abundance into a single diversity index using matrix analogues of Shannon's and Simpson's indices. *Ecography*, 2006; 29:525-530.
- [47]. Buzas MA, Hayek LAC. Biodiversity resolution: An integrated approach. *Biodiversity Letters*, 1996; 3:40-43.
- [48]. Omernik JM. The misuse of hydrologic unit maps for extrapolation, reporting, and ecosystem management. *Journal of America and Water Resources Assessment*, (2003; 39(3):563-573.
- [49]. Liu ZF, Liu GH, Fu BJ, Zheng XX. Relationship between plant species diversity and soil microbial functional diversity along a longitudinal gradient in temperate grasslands of Hulunbeir, Inner Mongolia, *China Ecological Resources*, 2008; 23:511–518.
- [50]. Melo AS. What we win "confounding" species richness and evenness in a diversity index? Biota Neotropica, 2008; 8: 021-027.
- [51]. Hayek LC, Buzas MA. Surveying Natural Populations. Columbia University Press, New York, 1997; Pp. 34-36.
- [52]. Purvis A, Hector A. Getting the measure of biodiversity. Nature, 2000; 405: 212-219.
- [53]. Baba MD, Dabai JS, Sanchi ID, Sabo AY. Profitability of Traditional Honey Production in Zuru Emirate, Kebbi State, Nigeria. World Rural Observations, 2014; 6(3): 44-49, <u>http://www.sciencepub.net/rural.</u>
- [54]. Girma SA. Agro-climatology of Millet Production in Desert Fringe Zone of Nigeria, A Case Study of Kebbi State. Unpublished M. Sc. dissertation. Federal University of Technology Minna, Niger state, 2008; Pp. 97.
- [55]. Ovington AJ, Haitkamp D, Lawrence DB, Jan N. Plant Biomass and Productivity of Prairie, Savannah, Oakwood, and Maize. Field Ecosystems in Central Minnesota, 2012; 44(1): 52–63.
- [56]. Jain SK, Rao RR. A handbook of field and herbarium methods. Today and Tomorrow's Printers and Publishers, New Delhi, 1977; Pp. 23-27.
- [57]. Ivens GW. Recent experiments on chemical control of *Eupatorium odoratum L*. Proceedings of the Third Nigerian Weed Science Group Meeting, 1973; pp 23–27.
- [58]. Hutchinson J, Dalziel JM. Flora of West Tropical Africa. Volumes 1 and 2. The White Friars Press Ltd, London, 1954; Pp. 34-69.
- [59]. Akobundu IO, Agyakwa CW. A handbook of West African weeds. IITA, Ibadan, Nigeria, 1998; Pp. 23-58.
- [60]. Fattebert J, Baubet E, Slotow R, Fischer C. Landscape effects on wild boar home range size under contrasting harvest regimes in a human-dominated agro-ecosystem. *European Journal of Wildlife Resources*, 2017; 63(2):32
- [61]. Harpole WS, Tilman D. Grassland Species Loss resulting from reduced niche dimension. Nature, 2007; 446: 791-793.
- [62]. Salisu N, Rabiu S. Soil chemical properties and plants species composition in Savanna ecosystem, North-western Nigeria. Savanna Journal of Basic and Applied Sciences, 2019; 1(1): 1-8.
- [63]. Youssef AM, Al-Fredan MA. Community Composition of major vegetation in the coastal area of Al-Uqair, Saudi Arabia in Response to ecological variations. *Journal of Biological Sciences*, 2008; 8: 713-721.
- [64]. Abd El-Wahab RH. Plant assemblage and diversity variation with human disturbances in coastal habitats of the western Arabian Gulf. *Journal of Arid Land*, 2016, 8, 787–798. [CrossRef]
- [65]. Neji M, Serbaji MM, Hardy O, Chaieb M. Floristic diversity and vegetation patterns along disturbance gradient in arid coasts in southern mediterranean: Case of the Gulf of Gabès, southern Tunisia. Arid Land Resource Management, 2018, 32, 291–315. [CrossRef]
- [66]. Nakahama N, Hirasawa Y, Minato T, Hasegawa M, Isagi Y, Shiga T. Recovery of genetic diversity in threatened plants through use of germinated seeds from herbarium specimens. *Plant Ecology*, 2015, 216, 1635–1647. [CrossRef]
- [67]. Das SKr, Ahmed RA, Sajan SK, Dash N, Sahoo P, Mohanta P, Sahu HK, Rout SD, Dutta SK. Diversity, distribution and species composition of odonates in buffer areas of Similipal Tiger reserve, Eastern Ghat, India. Academic Journal of Entomology, 2012; 5(1): 54-61, 2012.
- [68]. Bhandari BS, Nautiyal DC, Gaur RD. Structural attributes and productivity potential of an alpine pasture of Garhwal Himalaya. Journal of Indian Botanical Society, 1999; 78: 321-329.
- [69]. Wilsey B, Stirling G. Species richness and evenness respond in a different manner to propagule density in developing prairie microcosm communities. *Plant Ecology*, 2007; 190: 259–273
- [70]. Dash MC. Fundamental of Ecology. 2nd Edition Tata McGrawHill publishing company limited, New Delhi, 2003; Pp. 67-78.
- [71]. Javaid AS, Ashok K. Pandit. Application of diversity indices to crustacean community of Wular Lake, Kashmir Himalaya, *International Journal of Biodiversity and Conservation*, 2013; 5(6): 311-316, DOI: 10.5897/IJBC2013.0567