

Removal of Heavy Metals and Dyes Using Low Cost Adsorbents from Aqueous Medium-, A Review

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Abstract: The removal of heavy metals and dyes from waste water using low cost adsorbents is being widely used by various researchers. In recent years, the uses of various natural adsorbents have been widely used as a replacement for the current costly methods of treatment of waste water. In this review, an extensive list of recent investigations and literature has been compiled to provide information on a wide range of low cost adsorbents.

Key Words: Low cost adsorbents; Aqueous medium; Natural products; Micro organisms; Waste water.

I. Introduction:

Water pollution due to toxic heavy metals and dyes has been a major cause of concern for chemical and environmental engineers, which is because of excessive release of pollutants into the environment, due to industrialization and urbanization & poses a great problem to the environment.

As metals are nondegradable and dyes are highly complex, these two degrade very slowly. The waste water from industries like, metal plating, mining operations, tanneries, chloralkali and radiator manufacturing, smelting, alloy industries, textile industry, dye industries effluents need the treatment. The main techniques which have been utilized to reduce the heavy metal ions content and dyes of effluents include lime precipitation, ion exchange, adsorption onto activated carbon, membrane processing and electrolytic methods.

These methods have been found to be limited. Since they often involve high capital and operational costs and may be associated with the generation of secondary waste, which are the present treatment problems. But adsorption is very cheap, effective compared to other methods. The efforts have been contributed to develop new adsorbents and improving the existing adsorbents to have alternative materials to activated carbon.

Natural products as low cost adsorbents:

Srinivasan (1988) *et al.*, reported that, pH 2.5-3.0 was optimum with sulphuric acid treated rice husk for Chromium removal. Abraham (1998) *et al.*, studied Cadmium adsorption on Montmorillonite and reported at lower pH and with increasing concentration of humic acid, the metal ion does not adsorb. Mall and Upadhyay(1998) reported basic dyes adsorption on fly ash mainly depend on pH, increase in dye concentration, particle size, carbon content and agitation speed. Ajmal (1998) stated removal of Cadmium, Zinc, Nickel and Lead from aqueous solutions by *Mangifera Indica* which mainly depend on pH value and contact time and obeyed Freundlich adsorption isotherm. De and Basu(1999) reported that, increasing agitation time, carbon dosage and decreasing initial pH enhance the removal of direct-T, Blue-R from carbonized sugarcane bagasse. Baisakh and Patnaik (2002) stated, at ambient temperature condition, removal efficiency of chromium was more (70 %) by charcoal. Kannan & Meenakshi Sundaram (2002) reported that, Bamboo dust had higher adsorption capacity for dyes compared to Ground nut shell, Coconut shell, Rice husk and Straw. Amal raj and Srinivasa Raghavan (2002) and Suman Mor (2002) *et al.*, reported that, removal of Chromium was maximum at lower pH and at higher dose of activated carbon. Rao (2003) *et al.*, stated that, the removal of Cr⁶⁺, Ni²⁺, Cu²⁺, Pb²⁺ ions was more at pH 5.0-8.0. Katyal and Daga (2003) stated that, complete removal of dyes was found at pH 10 and 1.8 gm/L of adsorbent. Kannan and Umamathi (2003) reported that, the increase in dose of activated charcoal, contact time removal of Cd(II)-EDTA increases. Vasanth Kumar & Bagavanalu (2003), reported that, adsorption of dyes on Bioorganic waste shows the saturation point at 60 minutes and the rate kinetics was pseudo first order. Vasanthy (2003) *et al.*, stated that, mixture of adsorbents (Fly ash and activated carbon) removed about 90-100 % of Chromium at pH 2. Prabhakar & Palanivel (2003), reported orange TGU, removal depends on initial concentration of the dye and pH condition. Subanandam & Muttu Velagtydham (2003), stated that, adsorption of Methylene blue on lignite & fly ash increases with increase in adsorbent dosage, decreases in initial concentration, Shrihari and Raghavendra Kiran (2003), stated that, optimum value of contact time was 15 minutes and obeyed Freundlich isotherms for Iron adsorption. Sanjeev (2003) *et al.*, reported that, adsorption totally depends on dose, pH and concentration. Adsorption of dyes obeyed both Langmuir and Freundlich isotherms. Anima (2004) *et al.*, stated that, Ni (II) adsorption on rice husk depends on increase in time & dose. Kannan and Muthuraman (2004), stated that, dyes adsorption on activated charcoal increases with increase in contact time & dose.

Singh (2004) *et al.*, reported that, adsorption of copper on maize bran was at optimum pH 6.5, initial copper concentration 40.0 mg/l and temperature 20°C. Meena (2004) *et al.*, reported that, Copper adsorption on weathered coal sawdust and carbon aerogel was pH 6 dependent. Kannan and Balamurugan (2004), stated that, copper ions adsorption depends on dose of adsorbent and contact time. Jambulingam (2005) *et al.*, stated that, chromium ion adsorption depends on the initial concentration of metal ion, pH and agitation time. Srinivasan and Ramadevi (2006), stated that, lead adsorption on Tamarind nut carbon depends on adsorption dosage at an optimum pH of 5.0. Prabavathi Nagarajan and Priscilla Prabhavathi (2005), stated that, lead adsorption at pH 3, particle size 75µm and contact time 120 min was favourable for adsorption. Kannan and Ramamoorthy (2005) reported that, removal of dyes increased with contact time & dose. Senthil Nathan and Isaac Solomon Jebamani (2005), stated the Nickel (II) removal using wood of hybrid eucalyptus which depends on optimum dosage of 0.4g/ 150 ml and contact time was 10 minutes. Isaac Soloman Jebamani and Saseetharan (2005), reported that Nickel and Lead adsorption on Casuarinas equisetifolia wood was highly dependent on optimum pH 8.2, 20g/L and 15 minutes to remove 91.11 %.

Verma and Mishra (2005), reported that, dye removal was 87.5 % to 100 % from 20°C to 80°C, optimum was found to be 60°C. Kannan and Shakila (2005), reported that sky blue removal obeys Freundlich and Langmuir adsorption isotherms with first order rate kinetics and intraparticle diffusion at the rate determining step. Joshi and Srivastava (2005), stated Pipal bark after chemically carbonized adsorbs 76.04 %, at pH 2. Sharma *et al* (2005), reported that decolorising efficiencies of various adsorbents comparatively wood ash removes maximum color of Congored and Chrysofenine-G. Gharde *et al* (2006), stated that, Copper adsorption on Tamarindus Indica was pH dependent at a contact time of 90 minutes. Vasanthy and Sangeetha (2006), reported that, Chromium removal period was 4 hrs, with an adsorbent dosage of 1g/100 ml for 20 to 40 ppm, aqueous Chromium solution at pH 2, with intra-particle diffusion mechanism. Patil *et al* (2006), reported that Nickel removal by Babhul bark obeyed Langmuir and Freundlich models of adsorption. Mohammad Ajmal (2006), stated that, maximum adsorption of 85 % was observed at pH 6, data was correlated by Freundlich and Langmuir isotherm. Chaurasia and Shashikant (2007) stated that Langmuir and Freundlich models were found to have good agreement with experimental data for Methylene blue adsorption on activated saw dust, rice husk ash, boiler bottom ash and wood coal. Syed Shabudeen *et al.*(2006), stated that, temperature, dose and particle size had influence on adsorption.

Namasivayam and Sumitra (2006), stated that, waste Fe (III)/Cr (III) was used for Brilliant blue and Procion red removal and it was highly effective with agitation. Sheth and Umrigar (2006), stated that, Corncob carbon removed Cadmium through pore diffusion or intraparticle diffusion. Dongo Xinjiao (2006), stated that, Cladosporium species removed Copper at pH 5 and temperature 35°C and stirring speed of 100 rpm. Dipendu Banerjee (2007), stated that, fungi and bacteria removes iron at optimal temperature range 30°C, pH 2.5 and time 96 to 144 hrs. Mohapatra and Anand (2007), stated that, adsorption of Cd (II) on Lateritic ore obeyed Langmuir and Freundlich isotherms. The short contact time of 30 minutes and good loading capacity of 17.5 mg/g. Vijaya Kumari and Srinivasan (2007), stated that, Coconut oil cake residue removes mercury with wider pH range and dose. Lokeshappa *et al* (2007) stated that, Pseudomonas aeruginosa and Saccharomyces cerevasae found to remove Zinc and Nickel from aqueous solutions at death phase with 72 hr culture. Ahalya *et al.*(2007), reported that, Cajanus Cajan husk removes Chromium (VI) and Iron (III) and the adsorption was dependent on contact time, pH, initial metal ion concentration and biosorbent dosage. Padmini and Sridhar (2007) stated that Pongamia pinnata bark removes Manganese and Zinc with enhanced adsorbent dose at pH 7. Umesh (2007), stated that Ni (II) removal from sugarcane bagasse increases with contact time. Madhava Krishnan (2008) *et al.*, reported that, Ricinus communis pericarp carbon removes iron (II), with optimum pH, concentration and dose by obeying Langmuir and Freundlich isotherms. Raja Chandrasekar *et al.* (2008), stated that, iron adsorption from Borussus bark obeys Langmuir and Freundlich isotherms with positive R value and both the type of sorption (Chemisorption and Physisorption). Sneha Narverkar and Varsha (2008), stated that, Chromium sorption by Aspergillus Niger was pH dependent (2.0) Renganathan (2008), stated that, Congo Red dye removal by Tamarindus Indica fruit shell fitted very well with the Freundlich isotherm model. Kannan and Vijayakumar (2008), reported that, Ground nut shell and Coconut shell were correlated with Freundlich and Langmuir isotherms for the removal of Red industrial dye. Chandra sekhar (2008) stated that, removal of Lead by Coconut shell carbon obeyed first order reversible kinetic reaction and sorption isotherm followed Freundlich isotherm. Chakrapani *et al* (2008), stated that, Methylene blue by Maize shell carbon depends on pH 6.5, agitation time, dose of the adsorbent. Vasanthy *et al* (2008), stated that, Bacterial species adsorb Red RB dye by increasing the dose at acidic pH. Gandhimathi *et al* (2008), stated that, removal of Copper onto raw rice husk was enhanced at optimum contact time (90 min.) with a regression co-efficient of 0.999. Maji Sanjoy Kumar *et al* (2008), stated that, arsenic removal was dependent on the depth of the Laterite soil. Goyal Meenakshi *et al.*(2008), stated that, Mercury adsorption depends on carbon surface group, which involves attractive electrostatic interactions. Charumathi (2008) *et al.*, stated that, Lead (II) adsorption on pre-treated macro fungus obeyed Freundlich model. Mohammad Ajmal, *et al.*(2008), stated that, Lead adsorption on teak leaves depends on pH (5) and

temperature, with endothermic reaction. Sivamani and Prince Immanuel (2008), stated that, Pongamia pinnata removes chromium depending on pH and contact time. Dash and Satya Sagar (2008), stated that, Zinc adsorption on Shorea Robusta was totally depending on carbon dose, contact time and pH. Madhava Krishnan *et al.*(2008), stated that, Mercury (II) adsorption on Ricinus Communis carbon was dependent on pH, concentration and dose. Raju and Saseetharan (2008), stated that, Nickel (II) removal depends on optimum dosage, increase in contact time and pH 4.3. Kannan and Veemaraj (2009), stated that, Zinc removal by Jack fruit seed shows first order with intraparticle diffusion as one of the rate determining steps and depends on pH, contact time and dose.

Meenakshi Sundaram (2009), stated that, Azure-A dye removal from Pomegranate and Tamarind shell obeys Langmuir and Freundlich isotherms. Srinivasan and Hema (2009), stated that, Nickel (II) was adsorbed on Neem oil cake with increase in dose, contact time and pH. Gopalswami *et al* (2009), reported that, Methylene blue adsorption on Morus plant increased with contact time and achieved equilibrium within a short time. Kalpana *et al.* (2009), stated that, Terminalia Catappa removes Lead ions with pH 6, contact time 20 minutes, adsorption capacity 28.63 mg/g and correlation coefficient of 0.9996. Kale *et al.*(2009), stated decolorization and degradation of textile azo dye Golden Yellow HE2R was done to the extent of 90 % by bacteria. Varsha and Aditi (2009), stated that, white rot fungi *Trametes Versicolor* was used to remove Reactive black 5 as carbon source (glucose). Vijeta Gupta *et al.*(2009), stated Cadmium removal from modified Bagasse dust depend on pH 6.0, temperature 25°C, agitation time 5 hrs, sorbent dose 5 g/L and 150 rpm. Sonawane and Shrivastava (2009), stated that, Methylene blue adsorption on *Musa Paradisica* obeyed pseudo second order kinetics model. Sonawane and Shrivastava (2008), reported that, Malachite green adsorption on Banana leaves removed 100 % with appropriate dosage of the adsorbent. Pradeep Kumar *et al.*(2008), stated that, Nigrosine dye removal on petroleum based carbon depends on contact time and dose of adsorbent at pH 2. Latika Sharan *et al.*(2008), reported Cyanobacterium adsorption on Cadmium which depends on Contact time. Baskaran and Dhasekar (2008) stated that, removal of dyes by *Pleurotus Ostreatus* depends on pH 5-8 and it could be achieved by immobilizing the fungus with rice straw. Sudha and Celine (2008), stated Cadmium removal from chitisan coated coconut shell carbon was about 90 % with optimum contact time (120 minutes), agitation speed (60-80 rpm) in acidic condition. Santhalakshmi *et al.*(2008), stated that, chromium removal by sulphonated black rice husk and Sulphonated white rice husk ash was strongly dependent on pH 1.0, dose and contact time. Nour El. Din (2008), stated that, Chromium, Copper, Zinc, Cadmium and Lead removal by Nile Rose plant was dependent on pH and concentration. Suresh *et al.*(2009), reported that, Bentonite clays remove cationic species. The metal complex cations show better adsorption on Bentonite. Pravin *et al.* (2009), stated that, Red soil removes arsenic at pH 7.1 by obeying Langmuir and Freundlich adsorption isotherms. Raju and Saseetharan (2010), reported that, sludge based activated carbon removes Lead (II) at optimum contact time of 20 min, pH 5.3, with removal percentage 96. Kannan and Sarojini (2010), reported that, Manganese adsorption on commercial activated carbon depends on increase in contact time, dose of CAC and obey the Freundlich and Langmuir adsorption isotherms. Renugadevi *et al.*(2010), stated that, pods of wood apple removes Chromium (VI) with spontaneous and exothermic nature of adsorption. Meenakshi Sundaram *et al.*(2010), stated that, Nile blue A, Safranin and Ethyl violet on commercial activated carbon shows intraparticle adsorption and the process was first order. Nagashanmugam and Srinivasan (2010), reported that, Sulphuric and Zinc treated Gingelly oil cake removes Lead (II) at pH 5.0, dose of 0.1 g with 99.7 and 72.0 % respectively with pseudo-second order kinetics. The removal was 95.5 % with pH 4.0 and 0.1g by commercially activated carbon. Kannan and Veemaraj (2010), reported that, Cadmium and Cadmium- EDTA adsorb on Lemon peel carbon depending on metal ion concentration and pH dependent with First order intra-particle diffusion. Shrivastava (2010), stated that, Zinc (II) removal by granular activated carbon depends on pH 6 and removal increased with contact time. Renganathan *et al.*(2010), stated that, *Adathoda Vasica* removes maximum Chromium at pH 2 with sorbent dosage of 0.02g/100ml. The equilibrium data fit very well with Freundlich isotherm. Renugadevi *et al.* (2010), stated that, removal of methylene blue by *Cassia Siamea* depends on 180 min of contact time at pH 6 with 400 mg of dose. Sophie Beulah and Muthu Kumaran (2008), stated that, Chromium removal by *Syzygium Jambolanum* nut carbon was more when it was treated chemically, with the capacity of 4 times higher than CAC. Gupta Vikal and Sharma Manisha (2010), reported that, natural polymer Tamarind Kernal powder removes Chromium (VI) at 10 mg/L, with 65 % removal of Chromium (VI) at pH 5.0 and TKP dose level 5g/L. Yasmin Yamin *et al.*(2010), reported that, the percentage removal of amido black increased with increase in contact time and adsorbent dose by obeying Langmuir equation. Mallinga *et al.*(2010), stated that, human hair adsorption of Nickel (II) and Chromium (VI) were maximum at pH 6 and 4 respectively. Hence, the adsorption was totally pH dependent. Gupta Vikal *et al.* (2010), stated that, *Prosopis Aineraria* leaf powder removes Copper ions maximum in 240 minutes contact time and at increasing adsorbent amount, pH 5.5, fitting Langmuir and Pseudo second order equation. Ketcha Mbadcam Joseph and Bougo Tchamande Christelle (2010), stated that, Mercury (II) ions removed by Granular activated carbon and Kaolinite clay highly dependent on dose of adsorbent. The adsorption kinetics was pseudo first order.

Kannan et al.(2011), stated that, basic dyes onto teak leaf was totally dependent on contact time and dose of adsorbent at acidic pH by obeying first order kinetics with intraparticle diffusion. Kalaivani et al.(2011), stated that, adsorption of Reactive Red 2 and Acid blue 158 on to Chitin/ Chitisan was influenced by pH of the medium with spontaneous and endothermic in nature. Dilip Markandey et al. (2011), reported that, 144 hour old *Trichoderma Viridae* removes metals like Cadmium, Copper, Cobalt, Chromium, Nickel and Zinc from multimetallic aqueous solution at various pH ranges. Anantha Kumar and Krishnammal (2011), stated that, percentage removal of brilliant green on commercial activated charcoal increased with increase in contact time, dose and temperature with monolayer adsorption. Kannan and Veemaraj (2011), reported the Nickel (II) ions removal by Lemon peel and Pomegranate shell carbon, depending on particle size, initial concentration, contact time, dose and pH. Abdo Taher(2011), stated that, Methylene blue on Potato husk was favourable at pH 10, contact time 60 min, at thermodynamic values of $\Delta H=17.925$ KJ/mole and $\Delta S= 49.426$ J/ mol. Raman Kumar *et al.*(2011), stated that, Lead and Mercury can be removed by Fungal species which are supported by the presence of functional groups such as $-CH$, $C=O$ and $-OH$. Thamilarasu and Karunakaran (2011), stated that, Ni (II) removal by *Ricinus Communis* seed shell was at optimum contact time of 30 minutes, dosage 50 mg/50 ml by Freundlich adsorption. Dilip Markandey *et al.*(2011), stated that, Chromium (total) adsorb on *Rhizopus stonifer* depending on pH (1-3) and contact time with Chemisorption complexation reaction. Renugadevi *et al.*(2011), stated the malachite green removal using *Cassia fistula* which depends on pH 10 and 180 minutes as contact time, at initial concentration of 400 mg/L with 300 mg of the low-cost adsorbent. Theivarasu *et al.*(2011), stated the reactive orange 16 removal using cocoa shell carbon with monolayer adsorption capacity at 27.02 mg/g by using Langmuir Model equation with pseudo-second order kinetics equation. Pracilla Prabavathi *et al.*(2011), stated that, Rice husk and activated alumina were found to be good adsorbents, to remove Rhodamine B at pH 6 with maximum of 77% removal at pH 3. Kalpana *et al.*(2011), reported basic Violet 1 dye adsorption on *calotropis gigantean* biomass with uptake capacity of 138 mg/g at dose of 0.2 g/L, pH 2 and dye concentration of 110 mg/L. Mercy *et al.* (2011), stated that, *Terminalia chebula* removes Chromium with optimum metal ion concentration, adsorbent dose, agitation time and pH. Rosaline Vimla *et al.*(2011), stated the Mercury removal from Castor seed shell was maximum at pH 3 and 4 for CAC by obeying Freundlich isotherms. Renugadevi *et al.*(2011), reported Malachite green removal from *Caesalpinia Pulcherima* which depends on Contact time from 10 to 180 min, the adsorption increased with increase in pH 2-8 and 300 mg of the adsorption. Kannan and Kalimuthupandian (2011), stated Methylene blue removal using *Rutaceae Vila* carbon which depends on contact time and dose of adsorbent at basic pH. Malay Chaudhuri (2011), reported that, Red F-3B and Remazole Blue were removed by coconut coir carbon at pH 3.0, dose 10g/L and increased contact time to remove 96 % for Remazol Blue and 84 % for Remazol Red F-3B, obeying Langmuir and Freundlich adsorption isotherms.

Harminder Kaur *et al.* (2011), stated that *Kigelia Africana* was used to remove Copper (II) ions which obeyed Langmuir adsorption isotherm within 30 minutes with Maximum adsorption of 21.74 mg/g. Saritha Yadav et al. (2011), stated that, Congo-red dye removal by Rice husk which depends on initial dye concentration, increase in adsorbate-adsorbent contact period. Anusha and Suneeth Kumar (2011), stated iron adsorption using *Limonia acidissima* fruit shell which depends on pH 5.0, optimum dose and concentrations of 1.5g and 0.2 mg/L with contact time of 20 minutes respectively obeying Langmuir isotherm. Bhanupriya Mordhiya et al. (2012), reported the, Methylene blue adsorption on initial dye concentration, contact time, pH 7 obeying Freundlich and Langmuir isotherms with pseudo- second order model.

Based on this, the following adsorbents are used by the researchers for the removal of dyes and metal:

Table-1: Some of the low cost adsorbents used for the removal of dyes:

Dyes taken for adsorption studies	Low cost adsorbents used by investigators
Acid dye	Zinc activated bagasse ²⁰⁴ , Wood and Poots ^{134*}
Acid violet 17 (acid dye)	Orange peel ⁷³ , Orange peel ^{73&145}
Acid violet, Congo red and reactive orange	Banana pith ¹¹³
Acid orange-7 & direct red-31	Activated carbon ¹¹¹
Amido black dye	Calcinated and Uncalcinated Hydrotalcite ²¹⁷
Azure-A	CAC & Pomegranate shell and Tamarind shell ¹⁰⁴
Aniline Blue and Acid Violet	Commercially activated carbon (CAC) ⁷⁵
Basic violet-1	<i>Calotropis gigantea</i> ⁶⁴
Brilliant blue and Procion red	Fe(III)/Cr(III) ¹¹²
Brilliant green	Commercial Activated Carbon ¹¹

Congo red and Chrysofenine-G	(Wood charcoal, Saw dust, Wood ash, Brick powder, Fly ash, Sugarcane bagasse & Coir pith) ¹⁷¹ , Activated coir pith ¹¹⁴ , Tamarindus Indica fruit shell ⁵³ , Phanerochaetae chrysopodium ²⁰⁰ , Rice husk carbon ¹⁶⁵
Crystal violet	Pseudomonas putida ²⁶ , Fly ash and commercially Activated carbon ⁷¹ , Bacillus subtilis ²¹⁸ , Teak leaf ⁸⁰
Direct and acid dyes	Soya meal hull ¹⁴
Direct red and Acid brilliant blue	Banana pith ¹¹⁷
Direct T, Blue R,	Sugarcane bagasse ¹⁹⁵
Dye stuff effluent	Saw dust ²¹⁴
Direct blue	Tecomella Undulata (desert plant) ⁸³
Golden yellow HE2R	Adapted bacteria ⁶³
Malachite green	Banana ¹¹³ , Cassia fistula ¹⁵⁵ , Caesalpinia pulcherrima ¹⁵⁷ , Immobilised saccharomyces Cerevisiae ⁴³ , Banana leaves ¹⁸³ , Fly ash ⁹⁹
Methyl violet	Pseudomonas mendocina ¹⁶⁶
Methylene blue (Basic dye)	Rutaceae vila carbon ⁷⁰ , Cassia siamea ¹⁵⁴ , Cow dung ²⁰⁹ , (Rice husk ash, Boiler bottom ash, Saw dust ash & Wood coal) ²⁵ , Tea leaves ¹⁷⁸ , Oil palm fibre activated carbon ¹⁹⁹ , Jute fibre ¹⁶⁸ , Moringa oleifera ¹⁸ , Potato husk ¹ , Fly ash ⁹⁹ , Musa paradistica ¹⁸² , (Bamboo dust, Coconut shell, Ground nut shell, Rice husk and straw) ⁷¹ . Maize shell carbon ²² , Lignite fly ash ¹⁹³ , Saw dust ³² , Morus plant ⁴⁵ , ZnCl ₂ treated coir pith ¹¹⁵ , Wood ¹³⁵
Nigrosine dye	Petroleum based carbon ¹³⁸
Nile blue A, Safranin and Ethyl violet	Commercially activated carbon ¹⁰³
Orange II	White rot fungus (coriolus versicolor) ²¹⁹ , Tecomella Undulata (desert plant) ⁸³
Orange TGU	Palmnut shell and Fly ash ¹³⁷
Procion orangeM2R	Pseudomonas species S5 and Aspergillus flavus ¹⁷⁰
Procion red and Procion orange, Rhodamine B, Direct red12B, Congo red	Zn Cl ₂ treated coir pith ¹¹⁵
Reactive dye, Remazole Red, F-3B and Remazole blue.	Cashew nut hull carbon ²¹² , Coconut coir activated carbon ⁹⁹ .
Reactive orange and Basic Nile blue	Kopak hull activated carbon ¹⁹⁸
Reactive orange-16	Cocoa shell ²⁰³
Reactive red H8B and Direct green	Prosopis cineraria ¹⁶³
Reactive red 2 and acid blue	Chitin/ chitisan ⁶²
Red RB (reactive dye)	Enterobacter species, Escherichia coli and Bacillus ²¹¹
Reactive remzole red F-3B & Remzole blue	Coconut coir activated carbon ¹¹⁵
Rhodamine-B and Congo red	Orange peel ¹¹⁶
Rhodomine B and Reactive black 5	Rice husk and Activated alumina ¹⁴¹ , Fly ash & commercially activated carbon ¹⁰⁰ , Trametes versicolor (white rot fungi) ²⁰⁸ , Teak leaf ⁸⁰
Red industrial dye	Ground nut shell & Coconut shell carbon ⁷⁸
Selective textile dye effluent	Pleurotus ostreatus ¹⁷
Sky blue	Amla and coconut shell ⁷⁹
Victoria blue	Fly ash ⁸⁵
Xylenol Orange & Bromocresol purple, Acid orange-7 & Direct red-31	Activated charcoal (Carbon) ⁷²

* Number in superscript indicates the reference number in the references.

Some of the low cost adsorbents used for the removal of metals:

Metals	Adsorbents used
Arsenic	Laterite soil ⁹⁷ , Thiolated coconut fibre ⁵⁴ , Zirconium loaded activated charcoal ¹³¹ , Red soil ¹⁴⁰
Chromium	Rice husk ¹⁸⁹ , ZnCl ₂ treated coir pith ¹¹⁵ , Fly ash & activated carbon mixture ²¹³ , Groundnut husk ¹³⁰ , Coal char ¹⁶ , Prosopis spicigera ²¹⁰ , Cajanus cajan husk ³ , Aspergillus Niger ¹⁸¹ , Pongamia pinnata ¹²⁶ , Pipal bark ⁶¹ , Activated carbon ¹⁰ , Tamarind Kernel powder ⁴⁷ , Sulphonated black and White rice husk ¹⁶⁴ , Rhizopus stolonifer ³⁴ , Terminalia chebula ¹⁰⁵ , Adathoda Vasica ¹⁵² , Syzygium Jambolanum nut carbon ¹⁸⁴ , Pinus sylvestris bark ⁹ , Sorghum Vulgare (jowar) ¹⁷² , Simarouba glauca ¹¹⁹ , Bedi leaves ¹⁹¹ , Ipomea aquatica ⁷ , Hazelnut shell ⁸⁸ , Bone charcoal ²⁹ , Lignite coal ⁷⁶ , Tannin gel ²²⁰ , Waste tea leaves ¹⁷⁷ , Egg shells ²⁸ , Staphylococcus Saprophyticus ⁵⁶ , Oil-palm shell ¹⁴³ , Sugarcane leaves ¹²⁰ , Tobacco stem ¹⁷⁴ , Crescentia Cujeta (Thiruvottukai) ¹⁵⁸ , Ablesmoscus esculentus ⁶⁰ , Almond shell ²¹ , Corn cob ¹⁹ , Bagasse ¹⁵¹ , Human hair ¹⁰⁰ , Pods of wood apple ¹⁵⁶ , Activated Bagasse carbon ¹⁹⁶ , Parmelina tiliaceae (Lichen) ²⁰⁵ , Neem leaves ¹⁵ , Almond husk ⁵⁰ , Soya been hull ⁵⁹ , Trichoderma Viridae ³⁵ .
Cadmium	Lemon peel carbon ⁷⁷ , Lateritic ore ¹⁰⁸ , Nostoc carneum (cyanobacteria) ⁹¹ , Chitosan coated coconut ¹⁹⁴ , Montmorillonite ² , Corn cob (CCAC) ¹⁷³ , Ground nut husk ¹²⁵ , Wheat straw ³⁰ , Olive cake ¹⁶¹ , Botrytis cinerea fungal biomass ⁵ , Bone char ²⁷ , Modified ground nut husk ¹²⁵ , Fly ash & Sugar beet pulp ¹²⁹ , Fly ash & charcoal ⁸² , Spent grain ⁹⁴ , Jack fruit peel ¹⁹² , Cyanobacterium nostoc carneum agardh ⁹¹ , Orange waste ¹³² , Modified Dhoda bark ¹⁴⁹ , Mangifera indica ⁴ , Bagasse dust and Fly ash ²¹⁶ , Maize cob ¹²⁴ , Black gram husk ¹⁶² , Tea waste leaves ²⁰¹ , Almond husk ⁵⁰ , Trichoderma Viridae ³⁵
Cobalt	Maize cob ⁵³ , Trichoderma Viridae ³⁵
Copper	Kigelia Africana fruit carbon ⁴⁹ , Activated carbon (Bituminous coal origin) ¹⁶⁷ , Prosopis cineraria leaf powder ⁴⁸ , Tamarindus Indica ⁴¹ , Bagasse ¹⁵¹ , (Coal, Saw dust & Carbon aerogel) ¹⁰² , Fly ash ¹²⁷ , Sago waste ¹⁴² , Walnut shell ⁸⁶ , Activated carbon ³⁸ , Ulothrix zonata ¹²³ , Almond husk ⁵⁰ , Botrytis cinerea fungal biomass ⁵ , Streclulia futida fruit shell ¹⁸⁵ , Staphylococcus Saprophyticus ⁵⁶ , Peanut hull ¹³³ , Coconut husk powder ¹³⁹ , Tectona grandis ⁸⁷ , Maize cob ^{53,124} , Eucalyptus globules bark ³⁷ , Pulsa bark ¹⁵⁰ , Spent animal bones ⁸ , Cladosporium ³⁶ , Raw rice husk ⁴⁰ , Wheat straw ³⁰ , Oil palm ⁹³ , Coconut & Dates ⁶⁸ , Maize bran ¹⁷⁹ , Aegal Marmelore fruit shell ⁴² , Black gram ¹⁶² , Trichoderma Viridae ³⁵ ,
Gold	Activated Bagasse ²⁰⁷
Iron	Limonia acidissima (Wood apple) ¹³ , Thiobacillus ferroxidans and Bacteria ³³ , Cajanus cajan husk ³ , Ricinus communis pericarp carbon ⁹⁶ , Borassus bark ¹⁴⁴ , activated carbon ³⁸ , Rhizopus Arrhizus ⁶ , Maize cob ⁵³ , Laterite ¹⁷⁵ ,
Lead	Gingelly oil cake ¹⁰⁹ , Agaricus Bisporus ²⁴ , Coconut & Dates nut carbons ⁶⁹ , Casuarina Equisettifolia wood ⁵⁸ , Tamarind nut carbon ¹⁸⁷ , Tamarind seed and seed coat ¹³⁶ , Bagasse ¹⁵¹ , Teak leaves ¹⁰⁷ , Shorea Robusta ³¹ , Ground nut husk ¹²⁵ , Sago waste ¹⁴² , Ipomea aquatica ⁷ , Wheat bran ²⁰ , Olive cake ¹⁶¹ , Tea waste leaves ²⁰¹ , Tree fern ⁵² , Staphylococcus Saprophyticus ⁵⁶ , Spent grain ⁹⁴ , Phellinus Badius ¹⁰¹ , Lichen (Parmelina tiliaceae) ²⁰⁵ , Fruit stones ¹¹⁰ , Polymerised Banana stem ¹²¹ , Thiolated coconut fibre ⁵⁴ , Pre-treated spirulina biomass ⁴⁴ , Bamboo dust and Commercial activated carbons ⁸¹ , Pulsa bark ¹⁵⁰ , Coconut shell carbon ²³ , Mangifera Indica ⁴ , Terminalia catappa ⁶⁵ , Black gram husk ¹⁶² , Sludge based carbon ¹⁴⁷ , (Aspergillus flavus, Aspergillus nidulans, Phanerochaete chrysosporium, Penicillium Chrysogenum) ¹⁴⁸
Mercury	Castor seed shell ¹⁶⁰ , Activated carbons ⁴⁶ , ZnCl ₂ treated coir pith ¹¹⁵ , Ricinus Communis Pericarp ⁹⁵ , Thionated coconut fibre ⁵⁴ , Palm shell powder ⁸⁴ , Zirconium loaded activated charcoal ¹³¹ , Kaolinite clay and Granular activated carbon ⁸⁴ , Peanut hull ¹¹⁸ , Coconut oil cake residue ²¹⁵ , Terminalia catappa ⁵⁷ , (Aspergillus flavus, Aspergillus nidulans, Phanerochaete chrysosporium, Penicillium Chrysogenum) ¹⁴⁸ .
Molybdenum	ZnCl ₂ treated coir pith ¹¹⁵
Manganese	Pongamia pinnata bark ¹²⁶ , Commercially activated carbon ⁷⁴
Nickel	Sludge based activated carbon ¹⁴⁶ , Casuarina Equisettifolia wood ⁵⁸ , Activated alumina ¹⁵⁹ , Ground nut shell ¹⁰⁶ , Babul Bark & PAC ¹²⁸ , Sugar cane bagasse ²⁰⁶ , Pseudomonas aeruginosa and Saccharomyces cerevisiae ⁹² , Activated carbon ³⁸ , Coconut residues ¹⁸⁸ , Neem oil cake ¹⁸⁶ , Parthenium hysterophorus ⁹⁰ , sugarcane fibre ⁵⁵ , Eucalyptus globules bark ³⁷ , Aegal Marmelose fruit shell ⁴² , Pulsa bark ¹⁵⁰ , Rice husk ¹² , Bentonite clay ¹⁹⁷ , Mangifera Indica ⁴ , Human hair ¹⁰⁰ , Lemon peel carbon and Pomegranate shell ⁶⁷ , Spent animal bone ⁸ , Hybrid Eucalyptus ¹⁶⁹ , Almond husk ⁵¹ , Ricinus Communis seed shell ²⁰² ,

	Bagasse ¹⁵¹ , Trichoderma Viridae ³⁵
Selenium	Zirconium loaded activated charcoal ¹³¹ , ZnCl ₂ treated coir pith ¹¹⁵
Uranium	Pinus radiate ³⁹
Vanadium	ZnCl ₂ treated coir pith ¹¹⁵
Zinc	Tea leaves carbon ²⁰¹ , Sludge based activated carbon ¹⁴⁷ , Bituminous coal origin ¹⁶⁷ , Mangifera indica seed shell ⁴ , Pseudomonas aeruginosa and Saccharomyces cerevisiae ⁹² , Pongamia pinnata bark ¹²⁶ , Jack fruit seed & Commercial activated carbons ⁶⁶ , Fly ash and Sugar beet pulp ¹²⁹ , Palsa bark ¹⁵⁰ , Shorea Robusta (leaves) ³¹ , Fly ash ¹⁹⁰ , Sugarcane fibre ⁵⁵ , Trichoderma Viridae ³⁵ .

* Number in superscript indicates the reference number in the references.

II. Conclusion:

The use of commercially activated carbon can be replaced by the inexpensive and effective low cost adsorbents. There is need for more studies to understand better process of low-cost adsorbents and to demonstrate the technology effectively. Various low cost adsorbents show a high degree of removal efficiency for heavy metals and dyes. If low –cost adsorbents perform well in removing heavy metal complexes and dyes at low cost, they can be adopted and used widely in industries, not only to minimize cost but also to improve profit. In addition to this, the living organisms and the surrounding environment will also be benefited from the decrease or elimination of the potential toxicity due to the heavy metals and dyes

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