Gut Microbiota: The Missing Link in Obesity Induced Nonalcoholic Liver Disease

Roma Rattan¹, Purnendu Panda², Srikrushna Mahapatra³

¹Assistant Professor, Department of Biochemistry, S.C.B. Medical College, Cuttack ²Postgraduate Student, Department of Biochemistry, S.C.B. Medical College, Cuttack ³Professor and H.O.D, Department of Biochemistry, S.C.B. Medical College, Cuttack

Abstract

Background and objective: Recent research has elucidated a close association between intestinal microbiota, obesity, insulin resistance and nonalcoholic hepatic injury. Various studies have also indicated an increase in hepatic marker enzymes in obesity. Hence, this study aims to evaluate the association of gut microbiota with obesity, insulin resistance and hepatic marker enzymes.

Material and Method: This case-control study was conducted during March 2015 to October 2016 At S.C.B Medical College, Cuttack, Odisha. The study included 186 subjects (86 irritable bowel syndrome patients as per the Rome III criteria and hundred matched controls). Plasma fasting glucose, serum lipid profile, hepatic marker enzymes were analysed by commercial kits adapted to automated clinical chemistry analyser and serum fasting insulin was estimated by kits adapted to Lisa Scan.

Observation: Compared to controls the Irritable bowel syndrome patients had significantly higher Body mass index (20.9 ± 5.6 vs 30.1 ± 0.22), Waist-hip ratio (0.9 ± 0.11 vs 1.02 ± 0.06), lipid profile, hepatic marker enzymes and insulin resistance.

Conclusion: IBS patients were obese, and exhibited dyslipidemia, insulin resistance, elevated hepatic enzymes suggesting development of NAFLD.

Keywords: Irritable bowel syndrome, liver enzymes, obesity, nonalcoholic liver disease

I. Introduction

Recent research has revealed a close association between intestinal microbiota and host metabolic pathways (1-3). Various studies have revealed the association of intestinal microbiota with insulin resistance and obesity induced hepatic disease (4-5). The increase in intestinal microbiota cause irritable bowel syndrome (IBS) (6). IBS is illustrated by the following features- abdominal pain, discomfort, altered bowel habits without any detectable organic disease (7). IBS is commonly observed in the general population with a global prevalence of 7-30 % and its prevalence & incidence in India is 4 to 4.2% (8, 9, 10, 11). The pathophysiology of IBS is not clear and various hypotheses have been suggested, such as alterations in the intestinal microbiota, dysfunction of the gut-brain axis & autonomic nervous system, genetic predisposition, visceral antipathy, altered gastrointestinal hormones and obesity (12, 13). Various studies have indicated an association of increased intestinal bacteria with IBS and increased permeability of the intestine (14, 15). Intestinal permeability may be increased in the high fat diet of obesity (16). These metabolic alterations of increased microbiota and obesity lead to nonalcoholic fatty liver disease [NAFLD] (17, 18). Schnabl and Brenner suggested that the intestinal microbiota cause injury to the intestinal barrier and increase the permeability of bacterial endotoxins induce and help in progression of NAFLD (19). Hence, this study aims to evaluate the association of gut microbiota with obesity, insulin resistance and hepatic marker enzymes.

II. Material and Method

This case-control study was conducted during March 2015 to October 2016 At S.C.B Medical College, Cuttack, Odisha. The study included 186 subjects (86 irritable bowel syndrome patients as per the Rome III criteria and hundred matched controls). The IBS patients were further classified as per the Rome III criteria into IBS-C (constipation -predominent), IBS-D (diarrhea- predominent), IBS-M (mixed) and IBS-U (unsubtyped). As per the Rome III criteria, IBS is defined as recurrent abdominal discomfort or pain, at least for 3 days per month for 3 months with onset six months prior to diagnosis and associated with the following characteristics: (a) improvement with defecation, (b) onset associated with change in stool frequency, (c) onset associated with change in stool consistency (9, 20).

The anthropometric measurements were done while the study subjects were in light clothing and without shoes. Body mass index was calculated as weight in Kgs divided by height in meter². The waist circumference was assessed in the midway of the 12th rib and iliac crest. Blood pressure was measured after resting for 15 minutes. Venous blood samples were collected after overnight fast. Fasting blood sugar, lipid

profile, liver function test parameters were done by commercial kits adapted to Toshiba 120 FR autoanalyser. Serum fasting insulin was estimated by kits adapted to Lisa Scan. Insulin resistance was calculated by using the HOMA IR formula. HOMA index = fasting insulin (μ U/ml) X fasting blood sugar (mg/dl) /405. The HOMA index includes measurement of fasting plasma insulin and glucose and is calculated from these two parameters. The reference range of HOMA index is as follows:

> 2.0 indication for insulin resistance

> 2.5 insulin resistance probable

> 5.0 average value in patients with diabetes mellitus type 2

Statistical Analysis

All the data are represented as mean± standard deviation. An unpaired student's t test was done to compare the data. Pearson's correlation analysis was done to find the association. SPSS version 19 was used for all statistical analysis.

III. Results

The baseline demographic data of the study population is described in table 1. All the IBS patients were obese and had significantly higher BMI and waist hip ratio (WHR) as compared to the controls. Table 2 depicts the comparison of various biochemical parameters between the IBS patients and the control population. We observed a significant higher BMI and WHR in the IBS patients. We perceived statistically significant high serum total cholesterol, triglyceride, LDL cholesterol level in the IBS patients as compared to the heathy controls. The hepatic enzymes were significantly high in the IBS patients though Serum total bilirubin was within the normal range. The IBS patients were observed to have significant insulin resistance as shown by a higher HOMA IR level. Figure 1 shows the increase in hepatic enzymes

Table 1 Demographic data of the study population

Parameters	
Age in years	46 ± 5.4
Gender	male
BMI: IBS patients vs controls	30.1±0.22 vs 20.9 ±5.6*
Waist hip ratio: IBS patients vs controls	1.02±0.06 vs 0.9 ±0.11*
Systolic blood pressure mm of Hg	118 ± 2.66
Diastolic blood pressure mm of Hg	80 ± 6.66

Table 2 Comparison of biochemical parameters between the IBS patients and Controls

Parameters	IBS patients	Controls
Fasting blood sugar (mg%)	102 ± 1.22	98±3.1
Total cholesterol (mg%)	212±0.12	156± 0.56*
Triglycerides (mg%)	258±1.86	$104 \pm 1.28*$
HDL Cholesterol (mg%)	56.2 ± 0.89	49.4±0.66
LDL Cholesterol (mg%)	167 ± 1.46	111±2.4*
VLDL Cholesterol (mg%)	45.6 ± 0.24	20± 0.21
Total bilirubin (mg%)	0.5 ±0.02	0.5 ± 0.01
Alanine aminotransferase (IU/L)	42.1 ± 12.4	$24 \pm 10.2*$
Aspartate aminotransferase (IU/l)	39.9 ± 7.1	$20.1 \pm 5.4*$
Alkaline phosphatase (IU/l)	126.2 ± 14.2	$92.4 \pm 1.66*$
Gamma Glutamyl transferase (IU/L)	47.9 ± 1.4	$30.6 \pm 8.5*$
HOMA IR	2.2 ± 0.2	1.0 ±0.6*





IV. Discussion

We observed elevated hepatic enzymes in the IBS patients in this case control study. Our observation is similar to various previous studies (21, 22, and 23). Various studies have observed in the IBS, the intestinal microbiota is altered and there exists an overgrowth of the Gut microbiota (23). This causes increase release of bacterial endotoxins and alterations of the tight junctions leading to alterations in the intestinal permeability. The increased permeability leads to elevated bacterial endotoxins in the serum, which causes the release of proinflammatory molecules such as Tumor necrosis factor alpha and nitosative stress (19 - 18). These proinflammatory molecules and nitrosative stress induce hepatic injury which is observed by elevated hepatic enzymes.

Though few studies have examined the association of IBS and obesity, the pathophysiology is not completely elucidated. However, in vitro studies have suggested various mechanisms, which illustrate the lipid lowering role of intestinal microbiota, Bifidobacterium, in humans (29, 30). The end-products of bacterial fermentation, cholesterol assimilation, bile acid secretion help in lipid metabolism but in obesity alteration of intestinal microbiota affects lipid excretion (30). The endotoxins derived from intestinal bacteria link inflammation, insulin resistance to the high fat diet in obesity (29, 30). The bacterial endotoxins cause release of monocyte chemoattractant protein-1 which activates monocytes and releases various cytokines leading to inflammation, oxidative stress and insulin resistance (30). The dyslipidemia cause lipid deposition and NAFLD as seen by elevated hepatic enzymes.

In conclusion, we suggest estimation of bacterial endotoxins, in obese IBS patients and taking preparatory measures in reducing the alterations in intestinal bacteria as a preventive measure for NAFLD. **Conflict of interest** None

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